

Manuscript Number: THELANCETPSYCH-D-18-00678

Title: Multivariate meta-analyses of antidepressants, antipsychotics, anxiolytics, mood stabilizers, stimulants, antidementia and antiparkinsonian drug effects on mitochondrial complex I and IV in rodent models

Article Type: Article (Meta-analysis)

Keywords: meta-analysis; psychotropic drugs; NADH dehydrogenase; cytochrome-c-oxidase

Corresponding Author: Dr. Lisa Holper,

Corresponding Author's Institution: University Hospital of Psychiatry Zurich

First Author: Lisa Holper

Order of Authors: Lisa Holper; Dorit Ben-Shachar, Prof.; John J Mann, Prof.

Manuscript Region of Origin: SWITZERLAND

Abstract: Complex I (NADH dehydrogenase) and complex IV (cytochrome-c-oxidase) of the mitochondrial electron transport chain are reported to be affected by drugs used to treat psychiatric or neurodegenerative diseases, including antidepressants, antipsychotics, anxiolytics, mood stabilizers, stimulants, antidementia and antiparkinsonian drugs. We conducted meta-analyses examining the effects of each drug category on complex I and IV. The electronic databases Pubmed, EMBASE, CENTRAL and Google Scholar were searched for studies published between 1970 and 2018. Of 3105 screened studies, 68 articles covering 53 drugs were included in the meta-analyses. All studies assessed complex I and IV in rodent brain at the level of enzyme activity.

Meta-analyses revealed that selected antidepressants increase or decrease complex I and IV, antipsychotics and stimulants primarily decrease complex I but increase complex IV, whereas anxiolytics, mood stabilizers, antidementia and antiparkinsonian drugs preserve or even enhance both complex I and IV. To determine potential contributors to the drug effects, we meta-analyzed the drugs' neurotransmitter receptor profiles and found that affinity to adrenergic ( $\alpha$ 1B), dopaminergic (D1/2), glutaminergic (NMDA1,3), histaminergic (H1), muscarinic (M1,3), opioid (OP1-3), serotonergic (5-HT2A, 5-HT2C, 5-HT3A) and sigma ( $\sigma$ 1) receptors contributed most to the effects. We discuss the drug effects in relation to pharmacological mechanisms of action that might have relevance for clinical and research applications.

**Multivariate meta-analyses of antidepressants, antipsychotics, anxiolytics, mood stabilizers, stimulants, antimentia and antiparkinsonian drug effects on mitochondrial complex I and IV in rodent models**

Holper L <sup>1</sup>, Ben-Shachar D <sup>2</sup>, Mann JJ <sup>3</sup>

<sup>1</sup> Department of Psychiatry, Psychotherapy, and Psychosomatics, University Hospital of Psychiatry Zurich, University of Zurich, Switzerland

<sup>2</sup> Laboratory of Psychobiology, Department of Psychiatry, Rambam Health Care Campus, Rappaport Faculty of Medicine, Technion IIT, Haifa, Israel

<sup>3</sup> Division of Molecular Imaging and Neuropathology, Columbia University and New York State Psychiatric Institute, New York, USA

Corresponding author: Lisa Holper, Department of Psychiatry, Psychotherapy, and Psychosomatics, University Hospital of Psychiatry Zurich, University of Zurich, 8032 Zurich, Switzerland, Phone: +41-44-389 1576, Mail: [holper@ini.phys.ethz.ch](mailto:holper@ini.phys.ethz.ch)

## Summary

We conducted meta-analyses examining the effects of antidepressants, antipsychotics, anxiolytics, mood stabilizers, stimulants, antimentia and antiparkinsonian drugs on mitochondrial complex I and IV. The electronic databases Pubmed, EMBASE, CENTRAL, and Google Scholar were searched for studies published between 1970 and 2018. Results showed that antidepressants increase or decrease complex I and IV, antipsychotics and stimulants primarily decrease complex I but increase complex IV, whereas anxiolytics, mood stabilizers, antimentia, and antiparkinsonian drugs preserve or even enhance both complex I and IV. These effects could be related to the drugs' neurotransmitter receptor profiles. We discuss the drug effects in relation to pharmacological mechanisms of action that might have relevance for clinical and research applications.

## Abstract

**Background:** Complex I (NADH dehydrogenase) and complex IV (cytochrome-c-oxidase) of the mitochondrial electron transport chain are reported to be affected by drugs used to treat psychiatric or neurodegenerative diseases, including antidepressants, antipsychotics, anxiolytics, mood stabilizers, stimulants, antidementia, and antiparkinsonian drugs.

**Methods:** We conducted meta-analyses examining the effects of each drug category on complex I and IV. The electronic databases Pubmed, EMBASE, CENTRAL, and Google Scholar were searched for studies published between 1970 and 2018. Of 3105 screened studies, 68 articles covering 53 drugs were included in the meta-analyses. All studies assessed complex I and IV in rodent brain at the level of enzyme activity.

**Outcomes:** Meta-analyses revealed that selected antidepressants increase or decrease complex I and IV, antipsychotics and stimulants primarily decrease complex I but increase complex IV, whereas anxiolytics, mood stabilizers, antidementia, and antiparkinsonian drugs preserve or even enhance both complex I and IV. To determine potential contributors to the drug effects, we meta-analyzed the drugs' neurotransmitter receptor profiles and found that affinity to adrenergic ( $\alpha1B$ ), dopaminergic ( $D1/2$ ), glutaminergic (NMDA1,3), histaminergic ( $H1$ ), muscarinic ( $M1,3$ ), opioid ( $OP1-3$ ), serotonergic ( $5-HT_{2A}$ ,  $5-HT_{2C}$ ,  $5-HT_{3A}$ ) and sigma ( $\sigma1$ ) receptors contributed most to the effects.

**Interpretation:** We discuss the drug effects in relation to pharmacological mechanisms of action that might have relevance for clinical and research applications.

**Funding:** No specific funding.

**Keywords:** meta-analysis; psychotropic drugs; NADH dehydrogenase; cytochrome-c-oxidase.

## Research in context

**Evidence before this study:** Drugs used to treat psychiatric or neurodegenerative diseases, including antidepressants, antipsychotics, anxiolytics, mood stabilizers, stimulants, antimentia, and antiparkinsonian drugs are suggested to be one of the contributors to mitochondrial dysfunction observed in those disorders. Two enzymes have been shown to relate to mitochondrial function under those conditions, mitochondrial complex I (NADH dehydrogenase) and complex IV (cytochrome-c-oxidase). We aimed to meta-analyze the effects of each drug category on complex I and IV. We conducted a structured literature search in PubMed, EMBASE, CENTRAL, and Google Scholar to identify studies published between January 1970 and May 2018 using the search strings ‘NADH dehydrogenase’ OR ‘cytochrome-c-oxidase’ OR ‘complex I’ OR ‘complex IV’ AND ‘antidepressants’ OR ‘antipsychotics’ OR ‘anxiolytics’ OR ‘mood stabilizers’ OR ‘stimulants’ OR ‘Alzheimer’ OR ‘Parkinson’. Aggregated data were collected containing qualitative information (i.e., drug names, drug mechanisms of action, and brain regions of interest (ROIs)), quantitative data (i.e., number of drug-administered and control animals, dosage, and duration of drug administration), and outcomes (effect sizes in terms of standardized mean differences (SMD) and p-values).

**Added value of this study:** This meta-analysis summarizes the findings of drugs used to treat psychiatric and neurodegenerative disorders with regard to their effects on mitochondrial complex I and IV and relates those findings to the drugs’ neurotransmitter receptor profiles.

**Implications of all the available evidence:** Combined with existing evidence, the results of the meta-analysis may guide researchers and clinicians in the use of drugs to treat psychiatric and neurodegenerative disorders to consider how psychopharmacological agents and their neurotransmitter receptors profiles interact on mitochondrial function. Considering those relationships may help to reduce the burden of mitochondrial dysfunction in psychiatric and neurodegenerative conditions.

## Introduction

Impaired mitochondrial function is linked to the pathophysiology of major psychiatric disorders such as mood disorders and schizophrenia<sup>1-4</sup> and neurodegenerative disorders such as Alzheimer's and Parkinson's.<sup>5</sup> Mitochondrial function is essential for production of adenosine triphosphate (ATP), the main source of cellular energy. Impaired mitochondrial function results in decreased energy metabolism, reduced bioenergetics, oxidative stress, and apoptosis.<sup>3</sup>

Psychotropic medication used to treat psychiatric or neurodegenerative conditions may target mitochondrial dysfunction.<sup>6-8</sup> For instance, psychiatric drugs such as antidepressants are thought to inhibit mitochondrial respiration leading to decreased ATP production in animal models of depression.<sup>9</sup> Both typical and atypical antipsychotics impair mitochondrial function by inducing structural gene changes also implicated in depletion of ATP supply.<sup>10</sup> Anxiolytics such as benzodiazepines<sup>7</sup> and stimulants such as amphetamines<sup>11</sup> weaken mitochondrial function through the formation of free radicals, particularly reactive oxygen species. Conversely, mood stabilizers such as lithium or valproic acid have been shown to preserve or even enhance mitochondrial function by increasing the rate of cellular respiration.<sup>10</sup> Similarly, medication to treat neurodegenerative disorders, for example antidementia drugs such as cholinesterase inhibitors (ChEIs) or antiparkinsonian drugs such as the dopamine agonist levodopa<sup>12</sup> or monoamine oxidase inhibitors (MAOIs),<sup>13</sup> may have neuroprotective effects that restore and maintain mitochondrial function. Together, these effects may contribute to both adverse effects and efficacy of many psychotropic medications, not only in patients with mitochondrial disorders but also in the much wider population receiving these agents for psychiatric or neurodegenerative illnesses<sup>14</sup> as we have discussed in our previous meta-analysis.<sup>15</sup>

Prior to ATP generation, mitochondria direct electrons extracted from nutrients into a transmembrane proton gradient and this process is mediated by the electron transport chain (ETC). Two enzymes of the ETC located at the inner mitochondrial membrane are the most impaired in psychiatric and neurodegenerative disorders based on our recent meta-analysis<sup>15</sup> and the most affected ETC enzymes by psychotropic drugs.<sup>16</sup> The first enzyme, complex I (NADH dehydrogenase, NDU), is one of the entry enzymes of cellular respiration or oxidative phosphorylation in the mitochondrion. NDU is the largest multimeric ETC complex and a major contributor to the proton gradient across the

mitochondrial inner membrane, which drives ATP production. The second enzyme of interest, complex IV (cytochrome-c-oxidase, COX), catalyzes the final step in the ETC. Due to its rate-limiting role in this oxidative process<sup>17</sup> and its coupling with neuronal activation,<sup>18,19</sup> COX is proposed as a key marker of mitochondrial as well as neuronal function.<sup>20</sup> The two enzymes interact in that an assembled complex IV is required to maintain the stability of complex I<sup>21-23</sup>. Both complex I and IV also represent clinically relevant targets due to their accessibility using novel *in-vivo* technologies assessing complex I redox states<sup>24,25</sup> or oxidized complex IV.<sup>26</sup>

We chose the seven drug categories of antidepressants, antipsychotics, anxiolytics, mood stabilizers, stimulants, antimentia, and antiparkinsonian drugs, not only because of their potential common effects on mitochondrial function, but also based on the overlapping indications for which they are used to treat in clinical practice. Antidepressants are used in mood and anxiety disorders, but are also frequently applied to treat depressive symptomatology in schizophrenia and both Alzheimer's and Parkinson's.<sup>27,28</sup> Antipsychotics are used in schizophrenia as well as mood disorders to ameliorate psychotic episodes and used as antidepressants and anti-manic treatment.<sup>29</sup> Mood stabilizers are typically used for recurrent mood disorders but atypical antipsychotics can also be beneficial in schizophrenia due to their anti-aggressive effects<sup>30</sup> and possible improvement of negative symptoms.<sup>31</sup> Anxiolytics and stimulants are commonly given alone or in combination with antidepressants to treat comorbid anxiety,<sup>32</sup> fatigue, or attention deficits.<sup>33,34</sup> Antimentia drugs are not only used in Alzheimer's to improve cognitive function but also in Parkinson's<sup>35</sup> and schizophrenia<sup>36</sup> to treat comorbid dementia symptoms.

Besides the overlap in clinical application, the seven drug categories also show similarities in their mechanisms of action and neurotransmitter receptor and transporter binding profiles. Antidepressants, such as selective serotonin reuptake inhibitors (SSRIs), serotonin-norepinephrine reuptake inhibitors (SNRIs), or norepinephrine and dopamine reuptake inhibitors (NDRIs), primarily increase the levels of the monoamine neurotransmitters serotonin (5-hydroxytryptamine, 5-HT), norepinephrine, and dopamine (D) by inhibiting their reuptake at the corresponding transporters (SERT, NET, DAT);<sup>37-42</sup> to a lesser extent, some antidepressants also interact with adrenergic ( $\alpha,\beta$ ), glutaminergic ( $\alpha$ -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid, AMPA, N-methyl-D-aspartate, NMDA), histaminergic

(H), muscarinic (M), melatonergic (MT), and sigma ( $\sigma$ ) receptors. Typical antipsychotics exert their clinical effects as D2 receptor antagonists as their common target<sup>43,44</sup> and 5-HT<sub>2A</sub> receptor antagonists for atypical antipsychotics.<sup>45</sup> Occupancy of other 5-HT receptors and  $\alpha$ , H, M receptors contributes to side effects and adverse effects.<sup>46–48</sup> Anxiolytics such as benzodiazepines exert their various effects through allosteric modulation of gamma-aminobutyric acid (GABA-A) receptor.<sup>49</sup> Stimulants work by releasing dopamine and also have a moderate affinity for monoamine transporters (SERT, NET, DAT)<sup>50,51</sup> and to a lesser extent  $\alpha$  and D receptors.<sup>52</sup> Similarly, antidementia drugs such as cholinesterase inhibitors (ChEIs) and antiparkinsonian drugs such as the dopamine agonist levodopa target D receptors, inhibit acetylcholine (ACh) catabolism and have down-stream effects on 5-HT,  $\alpha$ , DAT and NMDA receptors.<sup>53</sup> These and other receptors have been shown to affect mitochondrial activities and vice versa.<sup>54,55</sup>

Numerous excellent reviews<sup>6,7,9–13</sup> have discussed the detailed effects on complex I and IV caused by each of the above-mentioned drug categories. A meta-analysis summarizing the findings across these drugs cannot be found in the literature. We conducted a literature review, screening for suitable papers and then conducted a meta-analysis examining the effects of each drug category on complex I and IV. This work extends our previous meta-analysis focusing on psychiatric and neurodegenerative disease levels.<sup>15</sup>



## **Materials and methods**

### **Literature search and study identification**

We conducted a structured literature search in PubMed, EMBASE, CENTRAL, and Google Scholar to identify studies published between January 1970 and May 2018 using the search strings ‘NADH dehydrogenase’ OR ‘cytochrome-c-oxidase’ OR ‘complex I’ OR ‘complex IV’ AND ‘antidepressants’ OR ‘antipsychotics’ OR ‘anxiolytics’ OR ‘mood stabilizers’ OR ‘stimulants’ OR ‘Alzheimer’ OR ‘Parkinson’. We additionally manually reviewed reference lists in all retrieved articles for related publications. Inclusion criteria were: studies published in the English language, studies investigated complex I and/or complex IV, studies reporting data in rodent disease models in comparison with a control group. Exclusion criteria were: studies in human patient populations, studies in other tissues than brain tissue, cellular studies, genetic studies, case reports, and publications not reporting original data.

### **Data extraction**

Aggregated data were extracted for each of the seven drugs categories: antidepressants, antipsychotics, anxiolytics, mood stabilizers, stimulants, antimentia, and antiparkinsonian drugs. Aggregated data contained qualitative information (i.e., drug names, drug mechanisms of action, and brain regions of interest (ROIs)), quantitative data (i.e., number of drug-administered and control animals, dosage, and duration of drug administration), and outcomes (effect sizes in terms of standardized mean differences (SMD) and p-values). In case of aggregated data not presented in the text, the authors were contacted for permission to reuse original data or data were read from figures. Mechanisms of action and receptor affinity profiles of each drugs were derived from DrugBank,<sup>56</sup> when available.

### **Multivariate random-effect meta-analyses**

Three separate meta-analyses were computed for complex I and IV separately to estimate: 1) the effects of drugs; 2) the effects of mechanisms of action; and 3) the effects of receptor affinity.

Moderators were built for each meta-analysis. ‘Drug’ moderators were based on the International Nonproprietary Names (INN) of the corresponding active pharmaceutical ingredients (e.g.,

‘escitalopram’ for citalopram). ‘Mechanism of action’ moderators were built by categorizing each drug according to its corresponding primary drug mechanism (e.g., ‘escitalopram’ was categorized as ‘SSRI’); drugs with unknown mechanisms of action were categorized according to their therapeutic uses (e.g., ‘valproic acid’ was categorized as ‘antiepileptic drug (AEG)’). ‘Receptor affinity’ moderators were built by assigning each drug to its corresponding affinity for one or more of the various neurotransmitter receptors, transporters, or enzymes,<sup>56</sup> if available. The type of receptor binding, i.e., agonism or antagonism (coded 1 and -1), was treated as a covariate. Not considered was binding affinity (i.e., the inhibitory constant,  $K_i$ ), since it is not available for all drugs.

All meta-analyses were performed using a multivariate, random-effect model based on the Metafor package<sup>57</sup> as implemented in R.<sup>58</sup> The multivariate model accounted for heterogeneity and dependency in the underlying true effects of multiple moderators that could overlap within subjects and studies (random factors). To adjust for dosage (mg/day) and administration duration (days) of the specific drugs, categorical dummy covariates were used. To adjust for sample size, effects were weighted based on study size. To allow for heterogeneity differences between moderators, an unstructured variance-covariance matrix was applied (function `rma.mv`; observed effects = SMD + dosage + duration; variance-covariance matrix = COV; weight = study size; variance structure = UN; moderators = ‘drugs’ OR ‘mechanism of action’ OR ‘receptor affinity : type’; random factors = subject + study; method = REML). A Wald-type test<sup>57</sup> was used to compare subgroup estimates between complex I and IV.

Heterogeneity was assessed using Cochran's Q-test and the inconsistency  $I^2$  statistic that directly indicates to what extent each outcome contributes to the total variance. Publication bias was assessed using Egger's regression analysis.

# Results

## Data extraction

Of 3105 screened studies, 68 studies were eligible for the meta-analyses (**Tab. 1, Dataset S1**) on antidepressants (AD), antipsychotics (AP), anxiolytics (AX), mood stabilizers (MS), stimulants (ST), antidementia drugs (ADD), and antiparkinsonian drugs (APD). In total, studies covered 53 drugs (AD N = 18, AP N = 10, AX N = 3, MS N = 5, ST N = 4, ADD N = 7, APD N = 7). The number of studies reporting on complex I (total N = 72, AD N = 24, AP N = 10, AX N = 4, MS N = 10, ST N = 9, ADD N = 6, APD N = 9) and complex IV (total N = 62, AD N = 26, AP N = 9, AX N = 3, MS N = 10, ST N = 6, ADD N = 5, APD N = 3) varied between drug categories, some of them reporting the same drug in more than one drug category. All studies analysed complex I and IV at the level of enzymatic activity in brain tissue from rodents (rats or mice) decapitated after drug administration. No eligible studies were found in healthy human brain tissue. Complex I and IV were analysed either in selected ROIs (AD N = 24, AP N = 7, AX N = 2, MS N = 9, ST N = 8, ADD N = 4, APD N = 7) and/or in brain homogenates (AD N = 14, AP N = 9, AX N = 2, MS N = 3, ST N = 5, ADD N = 4, APD N = 2). Drugs were investigated in one/two or more dosage steps (AD N = 18/10, AP N = 6/6, AX N = 3/1, MS N = 9/2, ST N = 5/4, ADD N = 4/2, APD N = 8/1). Mean (range) duration of drug administration was similar between drug categories (AD = 11 (1-28) days; AP = 17 (1-90) days; AX = 12 (1-30) days; MS = 14 (1-42) days; ST = 12 (1-28) days; ADD = 9 (1-21) days; APD = 5 (1-60) days). Mean number of animals per study (i.e., sum of drug-administered animals plus control animals) was also similar between drug categories (AD N = 14, AP N = 11, AX N = 21, MS N = 13, ST N = 13, ADD N = 15, APD N = 11). Control animals received saline or were sham groups in diseases models. Considering all above-mentioned factors, drug category, enzymes, selected ROIs, dosage, and administration duration, 1215 data points were extracted. Mechanisms of action and receptor affinity profiles collected based on previous work<sup>56</sup> are provided in **Tab. 1, Fig. S1, Dataset S2**.

## Multivariate random-effect meta-analyses

Meta-analyses assessed complex I and IV enzyme activity regarding the 1) effects of drugs (**Fig. 1**), 2) effects of mechanisms of action (**Fig. 2**), and 3) effects of receptor affinity (**Fig. 3**).

## Drugs

**Antidepressants** had negative effects on complex I for desipramine ( $p = 0.026$ ) and escitalopram ( $p < 0.0001$ ), whereas positive effects were observed for agomelatine ( $p = 0.00017$ ), paroxetine ( $p = 0.034$ ), and sertraline ( $p < 0.0001$ ) consistent with decreased versus increased NDU enzyme activity. Negative effects on complex IV were found for amitriptyline ( $p = 0.023$ ), desipramine ( $p < 0.0001$ ), fluoxetine ( $p = 0.045$ ), fluvoxamine ( $p < 0.0001$ ) and mirtazapine ( $p = 0.046$ ), whereas positive effects were observed for ketamine ( $p = 0.00021$ ), nortriptyline ( $p < 0.0001$ ), trazodone ( $p = 0.0014$ ), and sertraline ( $p = 0.023$ ). Differences in drug effects between complex I and IV were observed for agomelatine ( $p = 0.0003$ ), escitalopram ( $p < 0.0001$ ), paroxetine ( $p = 0.048$ ), and sertraline ( $p = 0.023$ ) as assessed using a Wild-type test.

**Antipsychotics** that had negative effects on complex I included chlorpromazine ( $p < 0.0001$ ), thiothixene ( $p = 0.00012$ ), and haloperidol ( $p < 0.0001$ ) consistent with decreased NDU enzyme activity. By contrast, positive effects on complex IV were observed for haloperidol ( $p = 0.039$ ) and risperidone ( $p = 0.00023$ ) indicating increased COX enzyme activity. All drug effects were significantly different between complex I and IV (chlorpromazine  $p < 0.0001$ , haloperidol  $p < 0.0001$ , risperidone  $p = 0.023$ , thiothixene  $p = 0.001$ ) as assessed using a Wild-type test.

**Anxiolytics** had a positive effect on complex IV for diazepam ( $p = 0.0014$ ), and no detectable effects were found on complex I.

**Mood stabilizers** had a positive effect on complex I for lithium ( $p = 0.02$ ) and on complex IV for gabapentin ( $p = 0.047$ ).

**Stimulants** having negative effects on complex I included amphetamine ( $p = 0.0099$ ), mazindol ( $p < 0.0001$ ), methamphetamine ( $p = 0.0012$ ), and methylphenidate ( $p < 0.0001$ ). There were no significant effects on complex IV.

**Antidementia drugs** having positive effects on complex IV included memantine ( $p = 0.034$ ) and latrepirdine ( $p = 0.029$ ). There were no significant effects on complex I.

**Antiparkinsonian drugs** having a positive effects on both complex I and complex IV were limited to selegiline ( $p = 0.013$  and  $p = 0.0033$ , respectively), without significant differences between complex I and IV effects.

### **Mechanisms of action**

Categorizing drugs according to their mechanisms of action revealed only few relevant findings compared to the abovementioned analysis. Negative effects on complex I were observed for typical antipsychotics ( $p < 0.0001$ ; Wild-type test  $p < 0.0001$ ), whereas lithium ( $p = 0.018$ ) and NDDIs ( $p = 0.01$ ; Wild-type test  $p = 0.025$ ) had positive effects. Positive effects on complex IV were also found for both AEGs ( $p = 0.046$ ) and NMDA antagonists ( $p = 0.0024$ ) (**Fig. 2**).

### **Receptor affinity**

Receptor affinity profiles were available for 45 drugs<sup>56</sup> (out of 56 drugs) (antidepressants  $N = 17$  (out of 18 drugs), antipsychotics  $N = 10$  (out of 10 drugs), anxiolytics  $N = 2$  (out of 3 drugs), mood stabilizers  $N = 4$  (out of 5 drugs), stimulants  $N = 4$  (out of 4 drugs), antidementia drugs  $N = 4$  (out of 7 drugs), antiparkinsonian drugs  $N = 4$  (out of 7 drugs)) (**Fig. S1, Dataset S2**). The results shown in **Fig. 3** thus represent information derived from combinations of drugs that share high affinity for a specific receptor but may differ in their affinity for other receptors.

For complex I, negative effects were found for 5-HT<sub>2A</sub> ( $p = 0.0044$ ),  $\alpha 1B$  ( $p = 0.027$ ), D1 ( $p = 0.00013$ ), D2 ( $p = 0.0021$ ), H1 ( $p = 0.022$ ), M1 ( $p = 0.017$ ), and M3 ( $p = 0.024$ ), whereas positive effects were observed for  $\sigma 1$  ( $p < 0.0001$ ). For complex IV, negative effects were found for 5-HT<sub>3A</sub> ( $p = 0.035$ ), DAT ( $p = 0.0035$ ), and NET ( $p = 0.014$ ), whereas positive effects were observed for 5-HT<sub>2C</sub> ( $p = 0.022$ ), NMDA1,3 ( $p = 0.00012$ ), OP1 ( $p = 0.0018$ ), OP2 ( $p = 0.0056$ ), OP3 ( $p = 0.016$ ), and  $\sigma 1$  ( $p = 0.021$ ). Differences in receptor effects on complex I and IV were found for 5-HT<sub>2A</sub> ( $p = 0.003$ ),  $\alpha 1A$  ( $p = 0.021$ ),  $\alpha 1B$  ( $p = 0.021$ ), D1 ( $p = 0.0002$ ), D2 ( $p = 0.014$ ), H1 ( $p = 0.026$ ), M1 ( $p = 0.045$ ), NMDA1,3 ( $p = 0.003$ ), and  $\sigma 1$  ( $p = 0.009$ ) as assessed using a Wild-type test.

None of abovementioned meta-analyses revealed relevant interaction effects with selected ROIs and we therefore do not report the corresponding results.

## Heterogeneity and publication bias

Overall, there was a high degree of heterogeneity as indicated by significant Q-statistics ( $p < 0.05$ ) and large  $I^2$  values (**Tab. 2**). Putative low heterogeneity was observed for stimulants in complex I ( $Q = 45.93$ ,  $p = 0.434$ ,  $I^2 = 11\%$ ), anxiolytics in complex IV ( $Q = 4.57$ ,  $p = 0.102$ ,  $I^2 = 82\%$ ), and antiparkinsonian drugs in complex IV ( $Q = 5.58$ ,  $p = 0.589$ ,  $I^2 = 34\%$ ), perhaps explained by the small study numbers in those drug categories (**Tab. 1**). Publication bias as assessed using Egger's regression test (**Tab. 2**) revealed non-significant results for all drug categories indicating no relevant publication bias.

## Discussion

This meta-analysis finds many drugs used to treat psychiatric and neurodegenerative disorders to affect mitochondrial complex I and IV. Overall antidepressants show the most heterogeneous effects on complex I and IV with some exhibiting negative effects while others have positive effects. Antipsychotics and stimulants primarily decrease complex I but increase complex IV. By contrast, anxiolytics, mood stabilizers, antidementia, and antiparkinsonian drugs affect both complex I and IV positively. To narrow down potential contributors to the drug effects, we meta-analysed mechanisms of action and receptor profiles and found that antagonism of adrenergic ( $\alpha_1B$ ), dopaminergic (D1,2), histaminergic (H1), muscarinic (M1,3) and serotonergic (5-HT<sub>2A</sub>) receptors result in negative effects on complex I, whereas agonism of sigma ( $\sigma_1$ ) receptors increases complex I. By contrast, antagonism of serotonergic (5-HT<sub>3A</sub>) receptors as well as inhibition of DAT and NET are associated with decreases in complex IV, whereas antagonism of glutaminergic (NMDA1,3) and serotonergic (5-HT<sub>2C</sub>) receptors as well as agonism of opioid (OP1-3) and sigma ( $\sigma_1$ ) receptors results in positive effects on complex IV. These findings support a relationship between drug effects and receptor affinity profiles and mitochondrial complex I and IV, which may have relevance for selection of treatment with low mitochondrial toxic potential in psychiatric and neurodegenerative conditions as we have discussed in our previous meta-analysis focusing on psychiatric and neurodegenerative disease levels.<sup>15</sup>

## Antidepressants

Antidepressants have heterogeneous effects on complex I and IV independent of their canonical mechanisms of action (**Fig. 1 & 2**). For example, while some SSRIs exhibit negative effects such as escitalopram on complex I<sup>59</sup> or fluoxetine and fluvoxamine on complex IV,<sup>60-64</sup> other SSRIs have positive effects such as paroxetine on complex I<sup>65,66</sup> and sertraline on both complex I and IV.<sup>67,68</sup> Similarly, some norepinephrine reuptake inhibitors (NRIs) such as desipramine and amitriptyline reveal strong negative effects on complex I and/or IV,<sup>16,64,69</sup> whereas nortriptyline, another NRI, affects complex IV positively.<sup>66</sup> Noradrenergic and specific serotonergic antidepressants (NaSSAs) such as mirtazapine have negative effects on complex IV (and I),<sup>16</sup> while serotonin antagonists and

reuptake inhibitors (SARIs) such as trazodone were observed to have a positive effect on complex IV.<sup>68</sup> The heterogeneity of these effects is likely related to the broader receptor profile of antidepressants such as antagonizing or agonizing adrenergic, dopaminergic, histaminergic, muscarinic, opioid, serotonergic and sigma receptors and inhibiting DAT, NET and SERT transporters. The combination of those receptor profiles in different antidepressants may lead to opposite effects on complex I and IV (**Fig. 3**). For example, antagonism of most serotonergic, i.e., 5-HT<sub>1A</sub>, 5-HT<sub>2A/C</sub>, 5-HT<sub>3</sub>, receptors has previously been shown to inhibit complex I, mitochondrial biogenesis and oxidative metabolism in rodent hippocampal neurons,<sup>70</sup> kidney,<sup>71,72</sup> and cardiac mitochondria,<sup>73</sup> respectively, in line with our findings (**Fig. 3**). Similarly, antagonism of muscarinic (M)<sup>74</sup> and histaminergic, especially H<sub>3</sub> brain-specific receptors,<sup>75</sup> inhibits natural mitochondrial protection from apoptotic and oxidative stress, the latter potentially antagonizing neuroprotective NMDA receptor activation.<sup>76</sup> Likewise, antagonism of adrenergic  $\alpha$ <sup>77</sup> and  $\beta$ <sup>78,79</sup> receptors can inhibit mitochondrial function by blocking their role in the stimulation of mitochondrial biogenesis (**Fig. 3**). By contrast, opioid receptors, especially delta-opioid receptors (OP1) (**Fig. 3**), are neuroprotective, particularly in cerebral cortex, by stabilizing ionic homeostasis, increasing antioxidant capacity and attenuating disrupted neuronal transmission.<sup>80,81</sup> Also, sigma ( $\sigma$ ) receptor agonism, especially  $\sigma$ 1 receptor (**Fig. 3**), which functions as a sensor for normal mitochondrial calcium (Ca<sup>2+</sup>) operation<sup>82</sup> and  $\sigma$ 1-ligands are thus discussed as protein-based pharmacological treatment to treat mitochondrial dysfunction in psychiatric and neurodegenerative conditions.<sup>83</sup>

Notably, agomelatine, a norepinephrine and dopamine disinhibitor (NDDI) and melatonergic MT<sub>1/2</sub> agonist, was found to have a prominent positive effect on complex I in this meta-analysis (**Fig. 1 & 2**). Previous work showed that low-dose agomelatine increases complex I activity in prefrontal cortex, cerebellum, and striatum, while decreasing it in the posterior cortex, whereas high-dose agomelatine decreases complex I activity in all those brain regions.<sup>84</sup> Agomelatine is thought to protect against pathology-induced decreases in complex I<sup>85-87</sup> by decreasing pathologically enhanced intracellular Ca<sup>2+</sup> levels<sup>88</sup> and inhibiting the opening of mitochondrial permeability transition pores.<sup>89</sup> This effect may be mediated by its agonist action at MT<sub>1</sub> and MT<sub>2</sub> receptors that have antioxidant activity and free radical scavenging, both actions are therapeutic for oxidative stress.<sup>90</sup>



Ketamine, a NMDA receptor antagonist, used as rapidly acting antidepressant,<sup>91</sup> was found to have a positive effect on complex IV (and to some extent complex I)<sup>92-95</sup> (**Fig. 1 & 2**). Previous work showed that ketamine increases complex IV but not complex I activity.<sup>93</sup> In the short-term (1 hour after treatment) it has been shown to increase complex IV activity in striatum and hippocampus but not in prefrontal cortex, whereas more delayed responses (after 6 hours of treatment) were described only in striatum.<sup>92</sup> Ketamine also increases other enzymes such as peroxiredoxins that enhance cellular antioxidant capacity by combating reactive oxygen species, reducing protein damage and adjusting homeostatic redox, which supports the role of mitochondria as downstream effectors mediating its rapid antidepressant action.<sup>96</sup> Ketamine's effects on complex IV may be reflected in the strong positive effects of the ionotropic glutamatergic NMDA receptor (**Fig. 3**) that is co-regulated by the same transcription factors as COX<sup>18,19</sup> and thought to restore the neuroprotective glutamatergic system.<sup>97</sup>

## Antipsychotics

Antipsychotics show strong negative effects on complex I primarily accounted for by the typical antipsychotics, chlorpromazine, thiothixene, and haloperidol<sup>98-104</sup> (**Fig. 1 & 2**). Antagonism of dopaminergic, primarily D2, receptors by typical antipsychotics (**Fig. 3**) may lead to dopamine accumulation in mitochondria resulting in D2-induced inhibition of complex I,<sup>105,106</sup> which in turn has been associated with the induction of extrapyramidal side effects (EPS).<sup>101,107</sup> By contrast, haloperidol affects complex IV positively<sup>102</sup> as supported by our analysis (**Fig. 1**). These opposing haloperidol effects on complex I and IV are thought to result from either up-stream (D2-induced) inhibition of complex I in the ETC<sup>21-23</sup> or reflect (D2-independent) enhanced functional neuronal activation due to the coupling between COX and neuronal activity.<sup>19</sup> This in turn may be clinically reflected in the delayed therapeutic effects of neuroleptic agents.<sup>102</sup>

By contrast, atypical antipsychotics were observed to have no significant effects besides an isolated increase in complex IV by risperidone<sup>65,98,99</sup> (**Fig. 1**), which might be related to their lower D2 receptor affinity for serotonergic 5-HT<sub>2A</sub>,<sup>108</sup> adrenergic, muscarinic, and histaminic receptors,<sup>44,109</sup> which also implies lower rates of EPS.

Together, these results support a complex I participation in dopamine-induced mitochondrial dysfunction such as observed in postmortem brain samples from deceased schizophrenic patients,<sup>110–115</sup> whereas the reduction in COX activity observed in schizophrenia<sup>15,116</sup> is thought to be independent of antipsychotic treatment.<sup>102</sup>

## Anxiolytics

Evidence that anxiolytics have weak positive effects on complex IV, is not certain because of the small number of studies in this drug category. Only the benzodiazepine, diazepam, a GABA-A receptor positive allosteric modulator (PAM) (**Fig. 2**), enhanced complex IV (**Fig. 1**), suggesting that this GABA-A effect may not be the mechanism because all the other benzodiazepines do not have this effect on complex IV. The possibility of an effect of GABA-A receptors (**Fig. 3**) was thus not supported by our analysis but needs to be further evaluated when more subjects have been studied because of the known neuroprotective effects of GABA-A agonists that may preserve COX activity by maintaining mitochondrial membrane potential, inhibiting downstream release of cytochrome c, and apoptotic signaling.<sup>117,118</sup>

## Mood stabilizers

Mood stabilizers show positive effects on both complex I and IV. Lithium, the most established mood stabilizer, increases complex I<sup>16,119–124</sup> (**Fig. 1 & 2**) by acting on intracellular Ca<sup>2+</sup> signaling, inhibiting inositol monophosphatase,<sup>4</sup> and modulating the neuroprotective ionotropic glutamatergic AMPA receptor,<sup>125–128</sup> which, however, did not reach significance in our analysis (**Fig. 3**). These findings are consistent with human studies in postmortem brain of patients with bipolar depression that report lithium stimulating mitochondrial complex I (but not complex IV) at clinically relevant concentrations.<sup>129,130</sup>

Antiepileptics, also used as mood stabilizers, such as gabapentin, enhance complex IV<sup>131,132</sup> (**Fig. 1 & 2**), which may be related to their NMDA receptors antagonism<sup>133</sup> since it showed positive effects for other drugs as well (**Fig. 3**). The main action of AEDs is to increase GABA transmission, which did show effects (**Fig. 3**). Although, little is known about the impact of AEDs on mitochondria, our findings suggest that some AEDs like gabapentin and lamotrigine) have beneficial effects despite

interfering with ETC-related membrane potential, mitochondrial biogenesis, morphology, dynamics, and survival.<sup>134</sup>

## **Stimulants**

Stimulants such as amphetamine, methamphetamine, mazindol and methylphenidate, all dopamine reuptake inhibitors (DRIs), affect complex I negatively<sup>135–143</sup> (**Fig. 1 & 2**) potentially related to their inhibition of DAT, NET and SERT transporters<sup>144</sup> (**Fig. 3**). Amphetamine enters and accumulates in mitochondria thereby dissipating the electrochemical gradient established by the ETC, inhibiting complex I and releasing cytochrome c, which in turn may induce the aforementioned interaction with complex IV and apoptosis.<sup>145</sup>

## **Antidementia drugs**

Antidementia drugs have no effects on complex I and it is not clear whether they affect complex IV (**Fig. 1 & 2**). Cholinesterase inhibitors (ChEIs), the most established antidementia drugs, inhibit ACh catabolism leading to its accumulation, greater stimulation of nicotinic and muscarinic receptors, and disrupted neurotransmission that may explain some of their positive effects on learning, memory, and other cognitive functions in Alzheimer's.<sup>146</sup> Memantine, a NMDA antagonist, also has a positive effect on complex IV<sup>147–149</sup> (**Fig. 1 & 2**) reflected in the strong positive NMDA receptor effect (**Fig. 3**) that is thought to restore unbalanced homeostasis in the glutamatergic system.<sup>150</sup> Positive effects on complex IV were also observed for latrepirdine<sup>149</sup> (**Fig. 1**); however, since its mechanism of action is not fully known its receptor affinity profile could not be assessed in the present analysis,<sup>56</sup> but may include neuroprotective effects by blocking neurotoxic Alzheimer beta-amyloid via modulation of NMDA receptors.<sup>151,152</sup> Together, these findings suggest that the well-documented deficit in complex IV in Alzheimer's,<sup>5,153,154</sup> thought to be due to neuronal toxicity and hypometabolism induced by beta-amyloid accumulation in mitochondria<sup>155–158</sup> can be addressed with antidementia drugs.

## **Antiparkinsonian drugs**

The most established antiparkinsonian drug, levodopa, an indirect dopamine agonist, was not found to affect complex I or IV.<sup>159–163</sup> This is in contrast to previous findings stating that levodopa significantly

increases complex I activity, while not changing complex IV activity<sup>161</sup> or that levodopa does not alter complex I activity by itself, but attenuates the decrease in complex I activity induced by MPTP (1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine), the prodrug to the neurotoxin MPP<sup>+</sup> (1-methyl-4-phenylpyridinium) which causes permanent symptoms of Parkinson's disease by destroying dopaminergic neurons in the substantia nigra.<sup>159</sup>

However, a weak positive effect on both complex I and IV was observed for selegiline, a selective irreversible MAO-B inhibitor in lower doses<sup>164</sup> (**Fig. 1 & 2**). Selegiline is thought to be neuroprotective by suppressing Ca<sup>2+</sup> efflux through mitochondrial permeability transition pores thereby inducing anti-apoptotic, pro-survival genes, though this is disputed.<sup>165</sup> Selegiline could thus protect mitochondrial function when used to treat Parkinson's symptoms in combination with levodopa,<sup>166</sup> but also off-label as palliative treatment for Alzheimer's dementia<sup>167</sup> and, as part of its antidepressant effects in major depressive disorder.<sup>168</sup>

## Methodological considerations

The present meta-analysis focused on rodent studies since no eligible studies were found in healthy human brain tissue. Doses and tissue levels of the cited drugs achieved in rodents are often much higher than those achieved in clinical treatment in humans. This is usually done to maximize effect but may create effects that are not clinically relevant. Such high doses may therefore have direct effects on mitochondrial electron transport, which may not be directly translatable to the effects expected at corresponding human doses. Further, we focused on drug effects at the neurotransmitter receptors level, but that is only one way drugs may indirectly alter mitochondrial activity. The observed considerable heterogeneity therefore needs to be considered indicating that not all effects can be explained at neurotransmitter receptors level. As an aside, the associations observed here are just "associations", they do not imply causality.

## Conclusion

Summarizing the overall strength of mitochondrial changes caused by the seven drug categories indicates that antidepressants have the most heterogeneous effects on both complex I and IV, which may be attributed to their broad receptor profiles including affinity to adrenergic ( $\alpha$ ), histaminergic

(H1-4), muscarinic (M1-5), opioid (OP1-3), serotonergic (5-HT<sub>2A</sub>, 5-HT<sub>2C</sub>, 5-HT<sub>3A</sub>), and sigma ( $\sigma$ 1) receptors. Typical versus atypical antipsychotics show strong negative effects on complex I versus weaker or no positive effects on complex IV, most likely associated with differential affinity for dopaminergic (D2), and serotonergic (5-HT<sub>2A</sub>) receptors. Stimulants also exhibit strong negative effects on complex I, potentially related to their inhibition of DAT, NET, and SERT transporters. Why stimulants and typical antipsychotics share a negative effect on complex I is hard to relate to the worsening of positive symptoms of psychosis by stimulants and the improvement by antipsychotics. In contrast, positive effects on complex I and/or IV are observed for anxiolytics, mood stabilizers, antidementia, and antiparkinsonian drugs, potentially associated with GABAergic (GABA-A), glutamatergic (AMPA, NMDA), and nicotinic (nACh) receptor effects. Further research is required, to better understand the connection between the actions of therapeutic agents and mitochondrial complexes. This may contribute to the development of mitochondrial targeted treatments in psychiatric and neurodegenerative disorders.<sup>169</sup>

## **Acknowledgements**

None.

## **Funding and Disclosure**

J.J.M. receives royalties for commercial use of the C-SSRS from the Research Foundation of Mental Hygiene. The remaining authors declare no competing interests.

## **Author contributions**

L.H. collected the data, performed the analysis, and drafted the manuscript. B.B.S. and J.J.M. provided helpful discussions and feedback to the manuscript. All authors agreed on the final version of the manuscript.

## **Data sharing statement**

Aggregated data collected for the meta-analysis are made available in the supplementary materials (Datasets 1 and 2).



## References

1. Manji H, Kato T, Di Prospero NA, Ness S, Beal MF, Krams M, et al. Impaired mitochondrial function in psychiatric disorders. *Nature Reviews Neuroscience* [Internet]. 2012 Apr 18;13:293. Available from: <http://dx.doi.org/10.1038/nrn3229>
2. Bansal Y, Kuhad A. Mitochondrial Dysfunction in Depression. *Current Neuropharmacology*. 2016;14(6):610–8.
3. Hroudová J, Fišar Z. Connectivity between mitochondrial functions and psychiatric disorders. *Psychiatry Clin Neurosci*. 2011;65(2):130–41.
4. Kato T. Neurobiological basis of bipolar disorder: Mitochondrial dysfunction hypothesis and beyond. *Schizophrenia Research*. 2017;187:62–6.
5. Onyango I, Khan S, Bennett J. Mitochondria in the pathophysiology of Alzheimer's and Parkinson's diseases. *Frontiers in Bioscience*. 2017;22:854–72.
6. Anglin R. Mitochondrial Dysfunction in Psychiatric Illness. *Canadian Journal of Psychiatry* *Revue Canadienne de Psychiatrie* [Internet]. 2016 Aug;61(8):444–5. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC4959647/>
7. Neustadt J, Pieczenik S. Medication-induced mitochondrial damage and disease. *Molecular Nutrition & Food Research* [Internet]. 2008 Jul 14 [cited 2018 May 28];52(7):780–8. Available from: <https://doi.org/10.1002/mnfr.200700075>
8. Ben-Shachar Dorit. Mitochondrial dysfunction in schizophrenia: a possible linkage to dopamine. *Journal of Neurochemistry* [Internet]. 2002 Dec 11 [cited 2018 Jul 3];83(6):1241–51. Available from: <https://doi.org/10.1046/j.1471-4159.2002.01263.x>
9. Adzic M, Brkic Z, Bulajic S, Mitic M, Radojicic MB. Antidepressant Action on Mitochondrial Dysfunction in Psychiatric Disorders. *Drug Dev Res* [Internet]. 2016 Nov 1;77(7):400–6. Available from: <http://dx.doi.org/10.1002/ddr.21332>
10. Manatt M, Chandra S. The effects of mitochondrial dysfunction in schizophrenia. Vol. 3. 2011. 84 p.
11. Brown JM, Yamamoto BK. Effects of amphetamines on mitochondrial function: role of free radicals and oxidative stress. *Pharmacology & Therapeutics* [Internet]. 2003 Jul 1;99(1):45–53. Available from: <http://www.sciencedirect.com/science/article/pii/S0163725803000524>
12. Winklhofer KF, Haass C. Mitochondrial dysfunction in Parkinson's disease. *Biochimica et Biophysica Acta (BBA) - Molecular Basis of Disease* [Internet]. 2010 Jan 1;1802(1):29–44. Available from: <http://www.sciencedirect.com/science/article/pii/S0925443909001963>
13. Hroudová J, Singh N, Fišar Z, Ghosh KK. Progress in drug development for Alzheimer's disease: An overview in relation to mitochondrial energy metabolism. *European Journal of Medicinal Chemistry* [Internet]. 2016 Oct 4;121:774–84. Available from: <http://www.sciencedirect.com/science/article/pii/S0223523416302665>
14. Anglin R, Rosebush P, Mazurek M. Psychotropic medications and mitochondrial toxicity. *Nature Reviews Neuroscience* [Internet]. 2012 Jul 25;13:650. Available from: <http://dx.doi.org/10.1038/nrn3229-c1>
15. Holper L, Ben-Shachar D, Mann J. Multivariate meta-analyses of mitochondrial complex I and IV in major depressive disorder, bipolar disorder, schizophrenia, Alzheimer disease, and Parkinson disease. *Neuropsychopharmacology* [Internet]. 2018 May 16; Available from: <https://doi.org/10.1038/s41386-018-0090-0>
16. Hroudová J, Fisar Z. Activities of respiratory chain complexes and citrate synthase influenced by pharmacologically different antidepressants and mood stabilizers. *Neuroendocrinology Letters*. 2010;31(3):336–42.
17. Arnold S. Cytochrome c Oxidase and Its Role in Neurodegeneration and Neuroprotection. In: Kadenbach B, editor. *Mitochondrial Oxidative Phosphorylation: Nuclear-Encoded Genes, Enzyme Regulation, and Pathophysiology* [Internet]. New York, NY: Springer New York; 2012. p. 305–39. Available from: [https://doi.org/10.1007/978-1-4614-3573-0\\_13](https://doi.org/10.1007/978-1-4614-3573-0_13)
18. Dhar SS, Wong-Riley MTT. Coupling of energy metabolism and synaptic transmission at the transcriptional level: Role of nuclear respiratory factor 1 in regulating both cytochrome c oxidase and NMDA glutamate receptor subunit genes. *The Journal of neuroscience : the official*

- journal of the Society for Neuroscience [Internet]. 2009 Jan 14;29(2):483–92. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC2775551/>
19. Wong-Riley MT. Bigenomic Regulation of Cytochrome c Oxidase in Neurons and the Tight Coupling Between Neuronal Activity and Energy Metabolism. *Advances in experimental medicine and biology* [Internet]. 2012;748:283–304. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC3710587/>
  20. Srinivasan S, Avadhani NG. Cytochrome c Oxidase Dysfunction in Oxidative Stress. *Free radical biology & medicine* [Internet]. 2012 Sep 15;53(6):1252–63. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC3436951/>
  21. Li Y, D'Aurelio M, Deng J-H, Park J-S, Manfredi G, Hu P, et al. An Assembled Complex IV Maintains the Stability and Activity of Complex I in Mammalian Mitochondria. *Journal of Biological Chemistry* [Internet]. 2007 Jun 15;282(24):17557–62. Available from: <http://www.jbc.org/content/282/24/17557.abstract>
  22. Diaz F, Fukui H, Garcia S, Moraes CT. Cytochrome c Oxidase Is Required for the Assembly/Stability of Respiratory Complex I in Mouse Fibroblasts. *Molecular and Cellular Biology* [Internet]. 2006 Jul;26(13):4872–81. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC1489173/>
  23. Schäfer E, Dencher NA, Vonck J, Parcej DN. Three-Dimensional Structure of the Respiratory Chain Supercomplex I<sub>1</sub>III<sub>2</sub>IV<sub>1</sub> from Bovine Heart Mitochondria,. *Biochemistry* [Internet]. 2007 Nov 1;46(44):12579–85. Available from: <https://doi.org/10.1021/bi700983h>
  24. Blacker TS, Duchen MR. Investigating mitochondrial redox state using NADH and NADPH autofluorescence. *Free Radical Biology & Medicine* [Internet]. 2016 Nov;100:53–65. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC5145803/>
  25. Zhu X-H, Lu M, Lee B-Y, Ugurbil K, Chen W. In vivo NAD assay reveals the intracellular NAD contents and redox state in healthy human brain and their age dependences. *Proceedings of the National Academy of Sciences of the United States of America* [Internet]. 2015 Mar 3;112(9):2876–81. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC4352772/>
  26. Bale G, Elwell C, Tachtsidis I. From Jöbsis to the Present Day: A Review of Clinical Near-Infrared Spectroscopy Measurements of Cerebral Cytochrome-C-Oxidase. *Journal of Biomedical Optics*. 2016;21(9):91307.
  27. Lanctôt KL, Amatniek J, Ancoli-Israel S, Arnold SE, Ballard C, Cohen-Mansfield J, et al. Neuropsychiatric signs and symptoms of Alzheimer's disease: New treatment paradigms. *Alzheimer's & Dementia: Translational Research & Clinical Interventions* [Internet]. 2017 Sep;3(3):440–9. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC5651439/>
  28. Zhuo C, Xue R, Luo L, Ji F, Tian H, Qu H, et al. Efficacy of antidepressive medication for depression in Parkinson disease: a network meta-analysis. *Tusconi. M, editor. Medicine* [Internet]. 2017 Jun;96(22):e6698. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC5459691/>
  29. Correll CU, Detraux J, De Lepeleire J, De Hert M. Effects of antipsychotics, antidepressants and mood stabilizers on risk for physical diseases in people with schizophrenia, depression and bipolar disorder. *World Psychiatry* [Internet]. 2015 Jun;14(2):119–36. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC4471960/>
  30. Fazel S, Zetterqvist J, Larsson H, Långström N, Lichtenstein P. Antipsychotics, mood stabilisers, and risk of violent crime. *Lancet* [Internet]. 2014 Sep 27;384(9949):1206–14. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC4165625/>
  31. Clelland CL, Drouet V, Rilett KC, Smeed JA, Nadrach RH, Rajparia A, et al. Evidence that COMT genotype and proline interact on negative-symptom outcomes in schizophrenia and bipolar disorder. *Translational Psychiatry* [Internet]. 2016 Sep 13;6:e891. Available from: <http://dx.doi.org/10.1038/tp.2016.157>
  32. Yu. Drobizhev M, V. Fedotova A, V. Kikta S. Antidepressants in Anxiety, Anxiolytics in Depression? Vol. 45. 2015. 413 p.
  33. Chen M-H, Pan T-L, Hsu J-W, Huang K-L, Su T-P, Li C-T, et al. Attention-deficit hyperactivity disorder comorbidity and antidepressant resistance among patients with major depression: A nationwide longitudinal study. *European Neuropsychopharmacology* [Internet]. 2016 Nov



- 1;26(11):1760–7. Available from: <http://www.sciencedirect.com/science/article/pii/S0924977X16305624>
34. Malhi GS, Byrow Y, Bassett D, Boyce P, Hopwood M, Lyndon W, et al. Stimulants for depression: On the up and up? *Aust N Z J Psychiatry* [Internet]. 2016 Feb 23 [cited 2018 Jun 23];50(3):203–7. Available from: <https://doi.org/10.1177/0004867416634208>
  35. O'Brien JT, Holmes C, Jones M, Jones R, Livingston G, McKeith I, et al. Clinical practice with anti-dementia drugs: A revised (third) consensus statement from the British Association for Psychopharmacology. *J Psychopharmacol* [Internet]. 2017 Jan 20 [cited 2018 May 28];31(2):147–68. Available from: <https://doi.org/10.1177/0269881116680924>
  36. Kishi T, Ikuta T, Oya K, Matsunaga S, Matsuda Y, Iwata N. Anti-Dementia Drugs for Psychopathology and Cognitive Impairment in Schizophrenia: A Systematic Review and Meta-Analysis. *International Journal of Neuropsychopharmacology* [Internet]. 2018 May 14;pyy045–pyy045. Available from: <http://dx.doi.org/10.1093/ijnp/pyy045>
  37. Murphy DL, Andrews AM, Wichems CH, Li Q, Tohda M, Greenberg B. Brain serotonin neurotransmission: An overview and update with an emphasis n serotonin subsystem heterogeneity, multiple receptors, interactions with other neurotransmitter systems, and consequent implications for understanding the actions of serotonergic drugs. *The Journal of Clinical Psychiatry*. 1998;59(Suppl 15):4–12.
  38. Tatsumi M, Groshan K, Blakely RD, Richelson E. Pharmacological profile of antidepressants and related compounds at human monoamine transporters. *European Journal of Pharmacology* [Internet]. 1997 Dec 11;340(2):249–58. Available from: <http://www.sciencedirect.com/science/article/pii/S0014299997013939>
  39. Owens MJ, Morgan WN, Plott SJ, Nemeroff CB. Neurotransmitter Receptor and Transporter Binding Profile of Antidepressants and Their Metabolites. *J Pharmacol Exp Ther* [Internet]. 1997 Dec 1;283(3):1305. Available from: <http://jpet.aspetjournals.org/content/283/3/1305.abstract>
  40. Cusack B, Nelson A, Richelson E. Binding of antidepressants to human brain receptors: focus on newer generation compounds. *Psychopharmacology*. 1994;114(4):559–65.
  41. Roth BL, Gibbons S, Arunotayanun W, Huang X-P, Setola V, Treble R, et al. The Ketamine Analogue Methoxetamine and 3- and 4-Methoxy Analogues of Phencyclidine Are High Affinity and Selective Ligands for the Glutamate NMDA Receptor. *PLOS ONE* [Internet]. 2013 Mar 19;8(3):e59334. Available from: <https://doi.org/10.1371/journal.pone.0059334>
  42. Hiranita T, J Kohut S, Soto P, Tanda G, Kopajtic T, Katz J. Preclinical Efficacy of N-Substituted Benzotropine Analogs as Antagonists of Methamphetamine Self-Administration in Rats. Vol. 348. 2013. 174 p.
  43. Correll CU. From receptor pharmacology to improved outcomes: individualising the selection, dosing, and switching of antipsychotics. *European Psychiatry* [Internet]. 2010 Jun 1 [cited 2018 May 21];25:S12–21. Available from: [http://dx.doi.org/10.1016/S0924-9338\(10\)71701-6](http://dx.doi.org/10.1016/S0924-9338(10)71701-6)
  44. Mauri M, Paletta S, Maffini M, Colasanti A, Dragogna F, Di Pace C, et al. Clinical pharmacology of atypical antipsychotics: an update. *EXCLI Journal* [Internet]. 2014;13:1163–91. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC4464358/>
  45. Sullivan LC, Clarke WP, Berg KA. Atypical Antipsychotics and Inverse Agonism at 5-HT(2) Receptors. *Current pharmaceutical design* [Internet]. 2015;21(26):3732–8. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC5543701/>
  46. Meltzer H, Massey B. The role of serotonin receptors in the action of atypical antipsychotic drugs. *Current Opinion in Pharmacology* [Internet]. 2011 Feb 1;11(1):59–67. Available from: <http://www.sciencedirect.com/science/article/pii/S1471489211000208>
  47. Scigliano G, Ronchetti G. Antipsychotic-Induced Metabolic and Cardiovascular Side Effects in Schizophrenia: A Novel Mechanistic Hypothesis. *CNS Drugs* [Internet]. 2013 Apr;27(4):249–57. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC3657088/>
  48. Humbert-Claude M, Davenas E, Gbahou F, Vincent L, Arrang J-M. Involvement of histamine receptors in the atypical antipsychotic profile of clozapine: a reassessment in vitro and in vivo. *Psychopharmacology* [Internet]. 2012 Mar 1;220(1):225–41. Available from: <https://doi.org/10.1007/s00213-011-2471-5>

49. Jensen H, Nichol K, Lee D, Ebert B. Clobazam and Its Active Metabolite N-desmethyloclobazam Display Significantly Greater Affinities for  $\alpha 2$ - versus  $\alpha 1$ -GABAA–Receptor Complexes. Vol. 9. 2014. e88456 p.
50. Han DD, Gu HH. Comparison of the monoamine transporters from human and mouse in their sensitivities to psychostimulant drugs. *BMC Pharmacology* [Internet]. 2006;6:6–6. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC1448202/>
51. Calipari ES, Ferris MJ. Amphetamine Mechanisms and Actions at the Dopamine Terminal Revisited. *The Journal of neuroscience : the official journal of the Society for Neuroscience* [Internet]. 2013 May 22;33(21):8923–5. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC3753078/>
52. BANERJEE SP, SHARMA VK, KUNG LS, CHANDA SK. Amphetamine induces  $\beta$ -adrenergic receptor supersensitivity. *Nature* [Internet]. 1978 Jan 26;271:380. Available from: <http://dx.doi.org/10.1038/271380a0>
53. Perez-Lloret S, Verónica Rey M, Crispo J, Krewski D, Lapeyre-Mestre M, Montastruc J-L, et al. Risk of heart failure following treatment with dopamine agonists in Parkinson’s disease patients. Vol. 13. 2014. 351 p.
54. Picard M, Wallace DC, Burelle Y. The rise of mitochondria in medicine. *Mitochondrion* [Internet]. 2016 Sep;30:105–16. Available from: <http://www.sciencedirect.com/science/article/pii/S1567724916300988>
55. Picard M, McEwen BS. Mitochondria impact brain function and cognition. *Proc Natl Acad Sci USA* [Internet]. 2014 Jan 7;111(1):7. Available from: <http://www.pnas.org/content/111/1/7.abstract>
56. Wishart DS, Feunang YD, Guo AC, Lo EJ, Marcu A, Grant JR, et al. DrugBank 5.0: a major update to the DrugBank database for 2018. *Nucleic Acids Research* [Internet]. 2018 Jan 4;46(Database issue):D1074–82. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC5753335/>
57. Viechtbauer W. Conducting Meta-Analyses in R with the metafor Package. *Journal of Statistical Software*. 2010;36(3):1–48.
58. R Development Core Team. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. 2008;
59. Gonçalves CL, Rezin GT, Ferreira GK, Jeremias IC, Cardoso MR, Carvalho-Silva M, et al. Differential effects of escitalopram administration on metabolic parameters of cortical and subcortical brain regions of Wistar rats. *Acta Neuropsychiatrica* [Internet]. 2012;24(3):147–54. Available from: <https://www.cambridge.org/core/article/differential-effects-of-escitalopram-administration-on-metabolic-parameters-of-cortical-and-subcortical-brain-regions-of-wistar-rats/8A4D5A973B301CFC771ACB182151F185>
60. Adzic M, Lukic I, Mitic M, Djordjevic J, Elaković I, Djordjevic A, et al. Brain region- and sex-specific modulation of mitochondrial glucocorticoid receptor phosphorylation in fluoxetine treated stressed rats: Effects on energy metabolism. *Psychoneuroendocrinology* [Internet]. 2013 Dec 1;38(12):2914–24. Available from: <http://www.sciencedirect.com/science/article/pii/S0306453013002692>
61. Ferreira GK, Cardoso MR, Jeremias I, Concalves C, Freitas K, Antonini R, et al. Fluvoxamine alters the activity of energy metabolism enzymes in the brain. *Revista Brasileira de Psiquiatria*. 2014;36(3).
62. Padilla E, Shumake J, Barrett DW, Sheridan EC, Gonzalez-Lima F. Mesolimbic effects of the antidepressant fluoxetine in Holtzman rats, a genetic strain with increased vulnerability to stress. *Brain research* [Internet]. 2011 Apr 28;1387:71–84. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC3081853/>
63. Shumake J, Colorado RA, Barrett DW, Gonzalez-Lima F. Metabolic mapping of the effects of the antidepressant fluoxetine on the brains of congenitally helpless rats. *Brain research* [Internet]. 2010 Jul 9;1343:218–25. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC2900439/>
64. Villa RF, Ferrari F, Bagini L, Gorini A, Brunello N, Tascedda F. Mitochondrial energy metabolism of rat hippocampus after treatment with the antidepressants desipramine and

- fluoxetine. *Neuropharmacology* [Internet]. 2017 Jul 15;121:30–8. Available from: <http://www.sciencedirect.com/science/article/pii/S0028390817301752>
65. Garabadu D, Ahmad A, Krishnamurthy S. Risperidone Attenuates Modified Stress–Re-stress Paradigm-Induced Mitochondrial Dysfunction and Apoptosis in Rats Exhibiting Post-traumatic Stress Disorder-Like Symptoms. *Journal of Molecular Neuroscience* [Internet]. 2015 Jun 1;56(2):299–312. Available from: <https://doi.org/10.1007/s12031-015-0532-7>
  66. Scaini G, Maggi DD, De-Nês BT, Gonçalves CL, Ferreira GK, Teodorak BP, et al. Activity of mitochondrial respiratory chain is increased by chronic administration of antidepressants. *Acta Neuropsychiatrica* [Internet]. 2011;23(3):112–8. Available from: <https://www.cambridge.org/core/article/activity-of-mitochondrial-respiratory-chain-is-increased-by-chronic-administration-of-antidepressants/8B69E10B16C0AF60CCC61E8D51D3FF65>
  67. Gaur V, Kumar A. Behavioral, biochemical and cellular correlates in the protective effect of sertraline against transient global ischemia induced behavioral despair: Possible involvement of nitric oxide-cyclic guanosine monophosphate study pathway. *Brain Research Bulletin* [Internet]. 2010 Apr 29;82(1):57–64. Available from: <http://www.sciencedirect.com/science/article/pii/S0361923010000298>
  68. Kumar P, Kalonia H, Kumar A. Nitric oxide mechanism in the protective effect of antidepressants against 3-nitropropionic acid-induced cognitive deficit, glutathione and mitochondrial alterations in animal model of Huntington’s disease. *Behavioural Pharmacology* [Internet]. 2010;21(3). Available from: [https://journals.lww.com/behaviouralpharm/Fulltext/2010/05000/Nitric\\_oxide\\_mechanism\\_in\\_the\\_protective\\_effect\\_of.6.aspx](https://journals.lww.com/behaviouralpharm/Fulltext/2010/05000/Nitric_oxide_mechanism_in_the_protective_effect_of.6.aspx)
  69. González-Pardo H, Conejo NM, Arias JL, Monleón S, Vinader-Caerols C, Parra A. Changes in brain oxidative metabolism induced by inhibitory avoidance learning and acute administration of amitriptyline. *Pharmacology Biochemistry and Behavior* [Internet]. 2008 May 1;89(3):456–62. Available from: <http://www.sciencedirect.com/science/article/pii/S0091305708000488>
  70. Chen S, Owens GC, Crossin KL, Edelman DB. Serotonin stimulates mitochondrial transport in hippocampal neurons. *Molecular and Cellular Neuroscience* [Internet]. 2007 Dec 1;36(4):472–83. Available from: <http://www.sciencedirect.com/science/article/pii/S1044743107001807>
  71. Harmon JL, Wills LP, McOmish CE, Demireva EY, Gingrich JA, Beeson CC, et al. 5-HT(2) Receptor Regulation of Mitochondrial Genes: Unexpected Pharmacological Effects of Agonists and Antagonists. *The Journal of Pharmacology and Experimental Therapeutics* [Internet]. 2016 Apr;357(1):1–9. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC4809314/>
  72. Rasbach KA, Funk JA, Jayavelu T, Green PT, Schnellmann RG. 5-Hydroxytryptamine Receptor Stimulation of Mitochondrial Biogenesis. *The Journal of Pharmacology and Experimental Therapeutics* [Internet]. 2010 Feb;332(2):632–9. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC2812119/>
  73. Wang Q, Zhang H, Xu H, Guo D, Shi H, Li Y, et al. 5-HTR3 and 5-HTR4 located on the mitochondrial membrane and functionally regulated mitochondrial functions. *Scientific Reports* [Internet]. 2016 Nov 22;6:37336. Available from: <http://dx.doi.org/10.1038/srep37336>
  74. De Sarno P, Shestopal SA, King TD, Zmijewska A, Song L, Jope RS. Muscarinic Receptor Activation Protects Cells from Apoptotic Effects of DNA Damage, Oxidative Stress, and Mitochondrial Inhibition. *The Journal of biological chemistry* [Internet]. 2003 Mar 28;278(13):11086–93. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC1361698/>
  75. Fernandez-Novoa L. Histamine and Immune Biomarkers in CNS Disorders. Vol. Volume 2016 (2016), Article ID 1924603, 10 pages. 2016.
  76. Hansen KB, Mullasseril P, Dawit S, Kurtkaya NL, Yuan H, Vance KM, et al. Implementation of a Fluorescence-Based Screening Assay Identifies Histamine H3 Receptor Antagonists Clobenpropit and Iodophenpropit as Subunit-Selective  $\alpha$ -Methyl-Aspartate Receptor Antagonists. *J Pharmacol Exp Ther* [Internet]. 2010 Jun 1;333(3):650. Available from: <http://jpet.aspetjournals.org/content/333/3/650.abstract>
  77. Beak JY, Huang W, Parker JS, Hicks ST, Patterson C, Simpson PC, et al. An Oral Selective Alpha-1A Adrenergic Receptor Agonist Prevents Doxorubicin Cardiotoxicity. *JACC: Basic to*

- Translational Science [Internet]. 2017 Feb 1;2(1):39–53. Available from: <http://www.sciencedirect.com/science/article/pii/S2452302X1630198X>
78. Cameron RB, Beeson CC, Schnellmann RG. Structural and pharmacological basis for the induction of mitochondrial biogenesis by formoterol but not clenbuterol. *Scientific Reports* [Internet]. 2017 Sep 5;7(1):10578. Available from: <https://doi.org/10.1038/s41598-017-11030-5>
  79. Peterson YK, Cameron RB, Wills LP, Trager RE, Lindsey CC, Beeson CC, et al.  $\beta(2)$ -adrenoceptor agonists in the regulation of mitochondrial biogenesis. *Bioorganic & medicinal chemistry letters* [Internet]. 2013 Oct 1;23(19):5376–81. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC3987705/>
  80. Guo J. Delta-Opioid Receptor-Mediated Protection and Mitochondria. In: Xia Y, editor. *Neural Functions of the Delta-Opioid Receptor* [Internet]. Cham: Springer International Publishing; 2015. p. 447–60. Available from: [https://doi.org/10.1007/978-3-319-25495-1\\_13](https://doi.org/10.1007/978-3-319-25495-1_13)
  81. He X, Sandhu HK, Yang Y, Hua F, Belser N, Kim DH, et al. Neuroprotection against hypoxia/ischemia:  $\delta$ -opioid receptor-mediated cellular/molecular events. *Cellular and Molecular Life Sciences* [Internet]. 2013 Jul 1;70(13):2291–303. Available from: <https://doi.org/10.1007/s00018-012-1167-2>
  82. Weng T-Y, Tsai S-YA, Su T-P. Roles of sigma-1 receptors on mitochondrial functions relevant to neurodegenerative diseases. *Journal of Biomedical Science* [Internet]. 2017;24:74. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC5603014/>
  83. Bolshakova AV, Kukanova EO, Gainullina AN, Zhemkov VA, Korban SA, Bezprozvanny IB. Sigma-1 receptor as a potential pharmacological target for the treatment of neuropathology. *St Petersburg Polytechnical University Journal: Physics and Mathematics* [Internet]. 2016 Mar 1;2(1):31–40. Available from: <http://www.sciencedirect.com/science/article/pii/S2405722316300214>
  84. de Mello A, Souza L, Cereja A, Schraiber E, Florentino D, Marins M, et al. Effect of subchronic administration of agomelatine on brain energy metabolism and oxidative stress parameters in rats. *Psychiatry and Clinical Neurosciences* [Internet]. 2015 Nov 9 [cited 2018 May 11];70(4):159–66. Available from: <https://doi.org/10.1111/pcn.12371>
  85. Gupta S, Sharma B. Pharmacological benefits of agomelatine and vanillin in experimental model of Huntington's disease. *Pharmacology Biochemistry and Behavior* [Internet]. 2014 Jul 1;122:122–35. Available from: <http://www.sciencedirect.com/science/article/pii/S0091305714001014>
  86. Kumar H, Sharma BM, Sharma B. Benefits of agomelatine in behavioral, neurochemical and blood brain barrier alterations in prenatal valproic acid induced autism spectrum disorder. *Neurochemistry International* [Internet]. 2015 Dec 1;91:34–45. Available from: <http://www.sciencedirect.com/science/article/pii/S0197018615300577>
  87. Singh P, Gupta S, Sharma B. Melatonin receptor and KATP channel modulation in experimental vascular dementia. *Physiology & Behavior* [Internet]. 2015 Apr 1;142:66–78. Available from: <http://www.sciencedirect.com/science/article/pii/S0031938415000736>
  88. Akpınar A, Uğuz AC, Nazıroğlu M. Agomelatine and Duloxetine Synergistically Modulates Apoptotic Pathway by Inhibiting Oxidative Stress Triggered Intracellular Calcium Entry in Neuronal PC12 Cells: Role of TRPM2 and Voltage-Gated Calcium Channels. *The Journal of Membrane Biology* [Internet]. 2014 May 1;247(5):451–9. Available from: <https://doi.org/10.1007/s00232-014-9652-1>
  89. Jia P, Liu C, Wu N, Jia D, Sun Y. Agomelatine protects against myocardial ischemia reperfusion injury by inhibiting mitochondrial permeability transition pore opening. *American Journal of Translational Research* [Internet]. 2018;10(5):1310–23. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC5992559/>
  90. Mahmood D, Muhammad BY, Alghani M, Anwar J, el-Lebban N, Haider M. Advancing role of melatonin in the treatment of neuropsychiatric disorders. *Egyptian Journal of Basic and Applied Sciences* [Internet]. 2016 Sep 1;3(3):203–18. Available from: <http://www.sciencedirect.com/science/article/pii/S2314808X16300197>

91. Duman R. Ketamine and rapid-acting antidepressants: a new era in the battle against depression and suicide [version 1; referees: 3 approved]. F1000Research [Internet]. 2018;7(659). Available from: <http://openr.es/c1j>
92. de Oliveira L, Fraga DB, De Luca RD, Canevar L, Ghedim FV, Matos MPP, et al. Behavioral changes and mitochondrial dysfunction in a rat model of schizophrenia induced by ketamine. *Metabolic Brain Disease* [Internet]. 2011 Mar 1;26(1):69–77. Available from: <https://doi.org/10.1007/s11011-011-9234-1>
93. Réus GZ, Nacif MP, Abelaira v M, Tomaz DB, Santos MAB dos, Anelise S. Carlessi, et al. Ketamine Treatment Partly Reverses Alterations in Brain Derived- Neurotrophic Factor, Oxidative Stress and Energy Metabolism Parameters Induced by an Animal Model of Depression. *Current Neurovascular Research* [Internet]. 2015;12(1):73–84. Available from: <http://www.eurekaselect.com/node/127943/article>
94. Rezin GT, Gonçalves CL, Daufenbach JF, Carvalho-Silva M, Borges LS, Vieira JS, et al. Effect of chronic administration of ketamine on the mitochondrial respiratory chain activity caused by chronic mild stress. *Acta Neuropsychiatrica* [Internet]. 2010;22(6):292–9. Available from: <https://www.cambridge.org/core/article/effect-of-chronic-administration-of-ketamine-on-the-mitochondrial-respiratory-chain-activity-caused-by-chronic-mild-stress/58E59E604E721E90828D757B732E5410>
95. Venâncio C, Félix L, Almeida V, Coutinho J, Antunes L, Peixoto F, et al. Acute Ketamine Impairs Mitochondrial Function and Promotes Superoxide Dismutase Activity in the Rat Brain. *Anesthesia & Analgesia* [Internet]. 2015;120(2). Available from: [https://journals.lww.com/anesthesia-analgesia/Fulltext/2015/02000/Acute\\_Ketamine\\_Impairs\\_Mitochondrial\\_Function\\_and.9.aspx](https://journals.lww.com/anesthesia-analgesia/Fulltext/2015/02000/Acute_Ketamine_Impairs_Mitochondrial_Function_and.9.aspx)
96. Weckmann K, Deery MJ, Howard JA, Feret R, Asara JM, Dethloff F, et al. Ketamine's antidepressant effect is mediated by energy metabolism and antioxidant defense system. *Scientific Reports* [Internet]. 2017;7:15788. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC5694011/>
97. Gough NR. Neuroprotective Mitochondrial Glutamate Receptors. *Sci Signal* [Internet]. 2012 Oct 23;5(247):ec272. Available from: <http://stke.sciencemag.org/content/5/247/ec272.abstract>
98. Balijepalli S, Boyd MR, Ravindranath V. Inhibition of mitochondrial complex I by haloperidol: the role of thiol oxidation. *Neuropharmacology* [Internet]. 1999 Apr 1;38(4):567–77. Available from: <http://www.sciencedirect.com/science/article/pii/S0028390898002159>
99. Balijepalli S, Kenchappa RS, Boyd MR, Ravindranath V. Protein thiol oxidation by haloperidol results in inhibition of mitochondrial complex I in brain regions: comparison with atypical antipsychotics. *Neurochemistry International* [Internet]. 2001 Apr 1;38(5):425–35. Available from: <http://www.sciencedirect.com/science/article/pii/S019701860000108X>
100. Barrientos A, Marín C, Miró O, Casademont J, Gómez M, Nunes V, et al. Biochemical and molecular effects of chronic haloperidol administration on brain and muscle mitochondria of rats. *Journal of Neuroscience Research* [Internet]. 1998 Dec 7 [cited 2018 May 13];53(4):475–81. Available from: [https://doi.org/10.1002/\(SICI\)1097-4547\(19980815\)53:4<475::AID-JNR9>3.0.CO;2-3](https://doi.org/10.1002/(SICI)1097-4547(19980815)53:4<475::AID-JNR9>3.0.CO;2-3)
101. Burkhardt C, Kelly P, Lim Y, Filley M, Parker D. Neuroleptic medications inhibit complex I of the electron transport chain. *Annals of Neurology* [Internet]. 1993 May [cited 2018 May 13];33(5):512–7. Available from: <https://doi.org/10.1002/ana.410330516>
102. Prince JA, Yassin MS, Orelan L. Neuroleptic-Induced Mitochondrial Enzyme Alterations in the Rat Brain. *J Pharmacol Exp Ther* [Internet]. 1997 Jan 1;280(1):261. Available from: <http://jpet.aspetjournals.org/content/280/1/261.abstract>
103. Prince JA, Yassin MS, Orelan L. A histochemical demonstration of altered cytochrome oxidase activity in the rat brain by neuroleptics. *European Neuropsychopharmacology* [Internet]. 1998 Feb 1;8(1):1–6. Available from: <http://www.sciencedirect.com/science/article/pii/S0924977X97000369>
104. Streck EL, Rezin GT, Barbosa LM, Assis LC, Grandi E, Quevedo J. Effect of antipsychotics on succinate dehydrogenase and cytochrome oxidase activities in rat brain. *Naunyn-Schmiedeberg's Archives of Pharmacology* [Internet]. 2007 Oct 1;376(1):127–33. Available from: <https://doi.org/10.1007/s00210-007-0178-2>

105. Ben-Shachar D, Zuk R, Gazawi H, Ljubuncic P. Dopamine toxicity involves mitochondrial complex I inhibition: implications to dopamine-related neuropsychiatric disorders. *Biochemical Pharmacology* [Internet]. 2004 May 15;67(10):1965–74. Available from: <http://www.sciencedirect.com/science/article/pii/S0006295204001303>
106. Brenner-Lavie H, Klein E, Zuk R, Gazawi H, Ljubuncic P, Ben-Shachar D. Dopamine modulates mitochondrial function in viable SH-SY5Y cells possibly via its interaction with complex I: Relevance to dopamine pathology in schizophrenia. *Biochimica et Biophysica Acta (BBA) - Bioenergetics* [Internet]. 2008 Feb 1;1777(2):173–85. Available from: <http://www.sciencedirect.com/science/article/pii/S0005272807002320>
107. Sykes DA, Moore H, Stott L, Holliday N, Javitch JA, Lane JR, et al. Extrapyramidal side effects of antipsychotics are linked to their association kinetics at dopamine D2 receptors. *Nature Communications* [Internet]. 2017 Oct 2;8(1):763. Available from: <https://doi.org/10.1038/s41467-017-00716-z>
108. Corena-McLeod M. Comparative Pharmacology of Risperidone and Paliperidone. *Drugs in R&D* [Internet]. 2015 Jun 1;15(2):163–74. Available from: <https://doi.org/10.1007/s40268-015-0092-x>
109. Seeman P. Atypical Antipsychotics: Mechanism of Action. *Can J Psychiatry* [Internet]. 2002 Feb 1 [cited 2018 Jun 15];47(1):29–40. Available from: <https://doi.org/10.1177/070674370204700106>
110. Ben-Shachar D, Zuk R, Gazawi H, Reshef A, Sheinkman A, Klein E. Increased mitochondrial complex I activity in platelets of schizophrenic patients. *International Journal of Neuropsychopharmacology* [Internet]. 1999 Dec 1;2(4):245–53. Available from: <http://dx.doi.org/10.1017/S1461145799001649>
111. Ben-Shachar D, Karry R. Neuroanatomical Pattern of Mitochondrial Complex I Pathology Varies between Schizophrenia, Bipolar Disorder and Major Depression. Hashimoto K, editor. *PLoS ONE* [Internet]. 2008;3(11):e3676. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC2579333/>
112. Dror N, Klein E, Karry R, Sheinkman A, Kirsh Z, Mazor M, et al. State-dependent alterations in mitochondrial complex I activity in platelets: a potential peripheral marker for schizophrenia. *Molecular Psychiatry* [Internet]. 2002 Oct 24;7:995. Available from: <http://dx.doi.org/10.1038/sj.mp.4001116>
113. Maurer I, Zierz S, Möller H-J. Evidence for a mitochondrial oxidative phosphorylation defect in brains from patients with schizophrenia. *Schizophrenia Research* [Internet]. 2001 Mar 1;48(1):125–36. Available from: <http://www.sciencedirect.com/science/article/pii/S092099640000075X>
114. Prince JA, Blennow K, Gottfries CG, Karlsson I, Oreland L. Mitochondrial Function is Differentially Altered in the Basal Ganglia of Chronic Schizophrenics. *Neuropsychopharmacology* [Internet]. 1999 Sep 1;21:372. Available from: [http://dx.doi.org/10.1016/S0893-133X\(99\)00016-0](http://dx.doi.org/10.1016/S0893-133X(99)00016-0)
115. Whatley S, Curti D, Marchbanks R. Mitochondrial involvement in schizophrenia and other functional psychoses. *Neurochemical Research*. 1996;21(9):995–1004.
116. Cavalier L, Jazin EE, Eriksson I, Prince J, Båve U, Oreland L, et al. Decreased Cytochrome-c Oxidase Activity and Lack of Age-Related Accumulation of Mitochondrial DNA Deletions in the Brains of Schizophrenics. *Genomics* [Internet]. 1995 Sep 1;29(1):217–24. Available from: <http://www.sciencedirect.com/science/article/pii/S0888754385712347>
117. Clarkson AN, Clarkson J, Jackson DM, Sammut IA. Mitochondrial involvement in transhemispheric diaschisis following hypoxia–ischemia: Clomethiazole-mediated amelioration. *Neuroscience* [Internet]. 2007 Jan 19;144(2):547–61. Available from: <http://www.sciencedirect.com/science/article/pii/S0306452206013042>
118. Tyagi N, Givvimani S, Kumar M, Kundu S, Gillespie WM, Mishra P, et al. Activation of GABA<sub>A</sub> receptor Protects Mitochondria and Reduces Cerebral ischemia. *The FASEB Journal* [Internet]. 2009 Apr 1 [cited 2018 Jun 15];23(1\_supplement):614.8–614.8. Available from: [https://www.fasebj.org/doi/abs/10.1096/fasebj.23.1\\_supplement.614.8](https://www.fasebj.org/doi/abs/10.1096/fasebj.23.1_supplement.614.8)
119. Bachmann RF, Wang Y, Yuan P, Zhou R, Li X, Alesci S, et al. Common effects of lithium and valproate on mitochondrial functions: protection against methamphetamine-induced

- mitochondrial damage. The international journal of neuropsychopharmacology / official scientific journal of the Collegium Internationale Neuropsychopharmacologicum (CINP) [Internet]. 2009 Jul;12(6):805–22. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC2779114/>
120. Feier G, Valvassori SS, Varela RB, Resende WR, Bavaresco DV, Morais MO, et al. Lithium and valproate modulate energy metabolism in an animal model of mania induced by methamphetamine. *Pharmacology Biochemistry and Behavior* [Internet]. 2013 Jan 1;103(3):589–96. Available from: <http://www.sciencedirect.com/science/article/pii/S0091305712002687>
  121. Kim HK, Isaacs-Trepanier C, Elmi N, Rapoport SI, Andreazza AC. Mitochondrial dysfunction and lipid peroxidation in rat frontal cortex by chronic NMDA administration can be partially prevented by lithium treatment. *Journal of psychiatric research* [Internet]. 2016 May;76:59–65. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC5843818/>
  122. Lambert PD, McGirr KM, Ely TD, Kilts CD, Kuhar MJ. Chronic Lithium Treatment Decreases Neuronal Activity in the Nucleus Accumbens and Cingulate Cortex of the Rat. *Neuropsychopharmacology* [Internet]. 1999 Aug 1;21:229. Available from: [http://dx.doi.org/10.1016/S0893-133X\(98\)00117-1](http://dx.doi.org/10.1016/S0893-133X(98)00117-1)
  123. Tan H, Young LT, Shao L, Che Y, Honer WG, Wang J-F. Mood stabilizer lithium inhibits amphetamine-increased 4-hydroxynonenal-protein adducts in rat frontal cortex. *International Journal of Neuropsychopharmacology* [Internet]. 2012 Oct 1;15(9):1275–85. Available from: <http://dx.doi.org/10.1017/S1461145711001416>
  124. Valvassori SS, Rezin GT, Ferreira CL, Moretti M, Gonçalves CL, Cardoso MR, et al. Effects of mood stabilizers on mitochondrial respiratory chain activity in brain of rats treated with d-amphetamine. *Journal of Psychiatric Research* [Internet]. 2010 Oct 1;44(14):903–9. Available from: <http://www.sciencedirect.com/science/article/pii/S0022395610000506>
  125. Du J, Gray A, Falke C, Yuan P, Szabo S, Manji K. Structurally Dissimilar Antimanic Agents Modulate Synaptic Plasticity by Regulating AMPA Glutamate Receptor Subunit GluR1 Synaptic Expression. *Annals of the New York Academy of Sciences* [Internet]. 2006 Jan 24 [cited 2018 Jun 22];1003(1):378–80. Available from: <https://doi.org/10.1196/annals.1300.031>
  126. Gould TD, O'Donnell KC, Dow ER, Du J, Chen G, Manji HK. Involvement of AMPA Receptors in the Antidepressant-Like Effects of Lithium in the Mouse Tail Suspension Test and Forced Swim Test. *Neuropharmacology* [Internet]. 2008 Mar;54(3):577–87. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC2275050/>
  127. Gray A, Du J, Falke C, Yuan P, Manji K. Lithium Regulates Total and Synaptic Expression of the AMPA Glutamate Receptor GluR2 in Vitro and in Vivo. *Annals of the New York Academy of Sciences* [Internet]. 2006 Jan 24 [cited 2018 Jun 22];1003(1):402–4. Available from: <https://doi.org/10.1196/annals.1300.036>
  128. Karkanas NB, Papke RL. Subtype-Specific Effects of Lithium on Glutamate Receptor Function. *Journal of Neurophysiology* [Internet]. 1999 Apr 1 [cited 2018 Jun 22];81(4):1506–12. Available from: <https://doi.org/10.1152/jn.1999.81.4.1506>
  129. Maurer IC, Schippel P, Volz H-P. Lithium-induced enhancement of mitochondrial oxidative phosphorylation in human brain tissue. *Bipolar Disorders* [Internet]. 2009 Aug 1;11(5):515–22. Available from: <http://dx.doi.org/10.1111/j.1399-5618.2009.00729.x>
  130. de Sousa RT, Streck EL, Zanetti MV, Ferreira GK, Diniz BS, Brunoni AR, et al. Lithium increases leukocyte mitochondrial complex I activity in bipolar disorder during depressive episodes. *Psychopharmacology* [Internet]. 2015 Jan 1;232(1):245–50. Available from: <https://doi.org/10.1007/s00213-014-3655-6>
  131. Kumar P, Kalonia H, Kumar A. Possible GABAergic mechanism in the neuroprotective effect of gabapentin and lamotrigine against 3-nitropropionic acid induced neurotoxicity. *European Journal of Pharmacology* [Internet]. 2012 Jan 15;674(2):265–74. Available from: <http://www.sciencedirect.com/science/article/pii/S001429991101497X>
  132. Kumar A, Lalitha S, Mishra J. Hesperidin potentiates the neuroprotective effects of diazepam and gabapentin against pentylenetetrazole-induced convulsions in mice: Possible behavioral, biochemical and mitochondrial alterations. *Indian Journal of Pharmacology* [Internet]. 2014;46(3):309–15. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC4071709/>

133. Chen J, Li L, Chen S-R, Chen H, Xie J-D, Sirrieh RE, et al. The  $\alpha\delta$ -1-NMDA Receptor Complex Is Critically Involved in Neuropathic Pain Development and Gabapentin Therapeutic Actions. *Cell reports* [Internet]. 2018 Feb 27;22(9):2307–21. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC5873963/>
134. Finsterer J, Scorza FA. Effects of antiepileptic drugs on mitochondrial functions, morphology, kinetics, biogenesis, and survival. *Epilepsy Research* [Internet]. 2017 Oct 1;136:5–11. Available from: <http://www.sciencedirect.com/science/article/pii/S0920121117302966>
135. Brown J, Quinton M, Yamamoto B. Methamphetamine-induced inhibition of mitochondrial complex II: roles of glutamate and peroxynitrite. *Journal of Neurochemistry* [Internet]. 2005 Sep 12 [cited 2018 May 17];95(2):429–36. Available from: <https://doi.org/10.1111/j.1471-4159.2005.03379.x>
136. Fagundes AO, Rezin GT, Zanette F, Grandi E, Assis LC, Dal-Pizzol F, et al. Chronic administration of methylphenidate activates mitochondrial respiratory chain in brain of young rats. *International Journal of Developmental Neuroscience* [Internet]. 2007 Feb 1;25(1):47–51. Available from: <http://www.sciencedirect.com/science/article/pii/S0736574806004448>
137. Fagundes AO, Aguiar MR, Aguiar CS, Scaini G, Sachet MU, Bernhardt NM, et al. Effect of Acute and Chronic Administration of Methylphenidate on Mitochondrial Respiratory Chain in the Brain of Young Rats. *Neurochemical Research* [Internet]. 2010 Nov 1;35(11):1675–80. Available from: <https://doi.org/10.1007/s11064-010-0229-9>
138. Gonçalves CL, Scaini G, Rezin GT, Jeremias IC, Bez GD, Daufenbach JF, et al. Effects of acute administration of mazindol on brain energy metabolism in adult mice. *Acta Neuropsychiatrica* [Internet]. 2014;26(3):146–54. Available from: <https://www.cambridge.org/core/article/effects-of-acute-administration-of-mazindol-on-brain-energy-metabolism-in-adult-mice/4C27C91BF0EFB09FF009F6FAE606DD69>
139. Killinger B, Shah M, Moszczynska A. Co-administration of betulinic acid and methamphetamine causes toxicity to dopaminergic and serotonergic nerve terminals in the striatum of late adolescent rats. *Journal of neurochemistry* [Internet]. 2014 Mar;128(5):764–75. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC4214679/>
140. Mishra J, Chaudhary T, Kumar A. Rosiglitazone Synergizes the Neuroprotective Effects of Valproic Acid Against Quinolinic Acid-Induced Neurotoxicity in Rats: Targeting PPAR $\gamma$  and HDAC Pathways. *Neurotoxicity Research* [Internet]. 2014 Aug 1;26(2):130–51. Available from: <https://doi.org/10.1007/s12640-014-9458-z>
141. Thrash B, Karuppagounder SS, Uthayathas S, Suppiramaniam V, Dhanasekaran M. Neurotoxic Effects of Methamphetamine. *Neurochemical Research* [Internet]. 2010 Jan 1;35(1):171–9. Available from: <https://doi.org/10.1007/s11064-009-0042-5>
142. Thrash-Williams B, Ahuja M, Karuppagounder SS, Uthayathas S, Suppiramaniam V, Dhanasekaran M. Assessment of Therapeutic Potential of Amantadine in Methamphetamine Induced Neurotoxicity. *Neurochemical Research* [Internet]. 2013 Oct 1;38(10):2084–94. Available from: <https://doi.org/10.1007/s11064-013-1117-x>
143. Thrash-Williams B, Karuppagounder SS, Bhattacharya D, Ahuja M, Suppiramaniam V, Dhanasekaran M. Methamphetamine-induced dopaminergic toxicity prevented owing to the neuroprotective effects of salicylic acid. *Life Sciences* [Internet]. 2016 Jun 1;154:24–9. Available from: <http://www.sciencedirect.com/science/article/pii/S0024320516301229>
144. Sitte HH, Freissmuth M. Amphetamines, new psychoactive drugs and the monoamine transporter cycle. *Trends in pharmacological sciences* [Internet]. 2015 Jan;36(1):41–50. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC4502921/>
145. Lowinson J, Ruiz P, Millmann R, Langrod J. Substance Abuse: A Comprehensive Textbook. New York, USA: Lippincott Williams & Wilkins; 2004. (P).
146. Čolović MB, Krstić DZ, Lazarević-Pašti TD, Bondžić AM, Vasić VM. Acetylcholinesterase Inhibitors: Pharmacology and Toxicology. *Current Neuropharmacology* [Internet]. 2013 May;11(3):315–35. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC3648782/>
147. Kumar H, Sharma B. Memantine ameliorates autistic behavior, biochemistry & blood brain barrier impairments in rats. *Brain Research Bulletin* [Internet]. 2016 Jun 1;124:27–39. Available from: <http://www.sciencedirect.com/science/article/pii/S0361923016300569>



148. Réus GZ, Stringari RB, Rezin GT, Fraga DB, Daufenbach JF, Scaini G, et al. Administration of memantine and imipramine alters mitochondrial respiratory chain and creatine kinase activities in rat brain. *Journal of Neural Transmission* [Internet]. 2012 Apr 1;119(4):481–91. Available from: <https://doi.org/10.1007/s00702-011-0718-2>
149. Singh N, Hroudová J, Fišar Z. In Vitro Effects of Cognitives and Nootropics on Mitochondrial Respiration and Monoamine Oxidase Activity. *Molecular Neurobiology* [Internet]. 2017 Oct 1;54(8):5894–904. Available from: <https://doi.org/10.1007/s12035-016-0121-y>
150. Parsons CG, Stöffler A, Danysz W. Memantine: a NMDA receptor antagonist that improves memory by restoration of homeostasis in the glutamatergic system - too little activation is bad, too much is even worse. *Neuropharmacology* [Internet]. 2007 Nov 1;53(6):699–723. Available from: <http://www.sciencedirect.com/science/article/pii/S0028390807002298>
151. Bachurin S, Shevtsova E, Kireeva E, Oxenkrug G, Sablin S. Mitochondria as a Target for Neurotoxins and Neuroprotective Agents. *Annals of the New York Academy of Sciences* [Internet]. 2006 Jan 24 [cited 2018 Jun 15];993(1):334–44. Available from: <https://doi.org/10.1111/j.1749-6632.2003.tb07541.x>
152. Grigor'ev VV, Dranyi OA, Bachurin SO. Comparative Study of Action Mechanisms of Dimebon and Memantine on AMPA- and NMDA-Subtypes Glutamate Receptors in Rat Cerebral Neurons. *Bulletin of Experimental Biology and Medicine* [Internet]. 2003 Nov 1;136(5):474–7. Available from: <https://doi.org/10.1023/B:BEBM.0000017097.75818.14>
153. Coskun P, Wyrembak J, Schriener S, Chen H-W, Marciniack C, LaFerla F, et al. A Mitochondrial Etiology of Alzheimer and Parkinson Disease. *Biochimica et Biophysica Acta* [Internet]. 2012 May;1820(5):553–64. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC3270155/>
154. Giachin G, Bouverot R, Acajjaoui S, Pantalone S, Soler-López M. Dynamics of Human Mitochondrial Complex I Assembly: Implications for Neurodegenerative Diseases. *Frontiers in Molecular Biosciences* [Internet]. 2016;3:43. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC4992684/>
155. Cenini G, Rüb C, Bruderek M, Voos W, Gilmore R. Amyloid  $\beta$ -peptides interfere with mitochondrial preprotein import competence by a coaggregation process. *MBoC* [Internet]. 2016 Sep 14 [cited 2018 Mar 31];27(21):3257–72. Available from: <https://www.molbiolcell.org/doi/abs/10.1091/mbc.e16-05-0313>
156. Picone P, Nuzzo D, Caruana L, Scafidi V, Di Carlo M. Mitochondrial Dysfunction: Different Routes to Alzheimer's Disease Therapy. *Oxidative Medicine and Cellular Longevity*. 2014;780179.
157. Pinho CM, Teixeira PF, Glaser E. Mitochondrial import and degradation of amyloid- $\beta$  peptide. *Biochimica et Biophysica Acta (BBA) - Bioenergetics* [Internet]. 2014 Jul 1;1837(7):1069–74. Available from: <http://www.sciencedirect.com/science/article/pii/S0005272814000541>
158. Readnower R, Sauerbeck A, Sullivan P. Mitochondria, Amyloid  $\beta$ , and Alzheimer's Disease. *International Journal of Alzheimer's Disease*. 2011;Article ID 104545.
159. Dixit A, Srivastava G, Verma D, Mishra M, Singh PK, Prakash O, et al. Minocycline, levodopa and MnTMPyP induced changes in the mitochondrial proteome profile of MPTP and maneb and paraquat mice models of Parkinson's disease. *Biochimica et Biophysica Acta (BBA) - Molecular Basis of Disease* [Internet]. 2013 Aug 1;1832(8):1227–40. Available from: <http://www.sciencedirect.com/science/article/pii/S0925443913001014>
160. Abdin AA, Hamouda HE. Mechanism of the neuroprotective role of coenzyme Q10 with or without L-dopa in rotenone-induced parkinsonism. *Neuropharmacology* [Internet]. 2008 Dec 1;55(8):1340–6. Available from: <http://www.sciencedirect.com/science/article/pii/S0028390808003821>
161. Calabrese Vittorio, Mancuso Cesare, Ravagna Agrippino, Perluigi Marzia, Cini Chiara, Marco Carlo De, et al. In vivo induction of heat shock proteins in the substantia nigra following L-DOPA administration is associated with increased activity of mitochondrial complex I and nitrosative stress in rats: regulation by glutathione redox state. *Journal of Neurochemistry* [Internet]. 2007 Jan 4 [cited 2018 May 20];101(3):709–17. Available from: <https://doi.org/10.1111/j.1471-4159.2006.04367.x>

162. Przedborski Serge, Jackson-Lewis Vernice, Muthane Uday, Jiang Hong, Ferreira Mayra, Naini Ali B., et al. Chronic levodopa administration alters cerebral mitochondrial respiratory chain activity. *Annals of Neurology* [Internet]. 1993 Nov [cited 2018 May 20];34(5):715–23. Available from: <https://doi.org/10.1002/ana.410340515>
163. Sharma N, Jamwal S, Kumar P. Beneficial effect of antidepressants against rotenone induced Parkinsonism like symptoms in rats. *Pathophysiology* [Internet]. 2016 Jun 1;23(2):123–34. Available from: <http://www.sciencedirect.com/science/article/pii/S0928468016300050>
164. Czerniczyniec A, Bustamante J, Lores-Arnaiz S. Modulation of brain mitochondrial function by deprenyl. *Neurochemistry International* [Internet]. 2006 Feb 1;48(3):235–41. Available from: <http://www.sciencedirect.com/science/article/pii/S0197018605002433>
165. Wu Y, Kazumura K, Maruyama W, Osawa T, Naoi M. Rasagiline and selegiline suppress calcium efflux from mitochondria by PK11195-induced opening of mitochondrial permeability transition pore: a novel anti-apoptotic function for neuroprotection. *Journal of Neural Transmission* [Internet]. 2015 Oct 1;122(10):1399–407. Available from: <https://doi.org/10.1007/s00702-015-1398-0>
166. Riederer P, Lachenmayer L, Laux G. Clinical Applications of MAO-Inhibitors. *Current Medicinal Chemistry* [Internet]. 2004;11(15):2033–43. Available from: <http://www.eurkaselect.com/node/62094/article>
167. Birks J, Flicker L. Selegiline for Alzheimer's disease. *Cochrane Database of Systematic Reviews* [Internet]. 2003;(1). Available from: <http://dx.doi.org/10.1002/14651858.CD000442>
168. Citrome L, Goldberg JF, Portland KB. Placing transdermal selegiline for major depressive disorder into clinical context: Number needed to treat, number needed to harm, and likelihood to be helped or harmed. *Journal of Affective Disorders* [Internet]. 2013 Nov 1 [cited 2018 Jun 16];151(2):409–17. Available from: <http://dx.doi.org/10.1016/j.jad.2013.06.027>
169. Ben-Shachar D, Ene HM. Mitochondrial Targeted Therapies: Where Do We Stand in Mental Disorders? *Biological Psychiatry* [Internet]. 2017 Aug 15; Available from: <http://www.sciencedirect.com/science/article/pii/S0006322317318590>
170. Singh A, Kumar A. Microglial Inhibitory Mechanism of Coenzyme Q10 Against A $\beta$  (1-42) Induced Cognitive Dysfunctions: Possible Behavioral, Biochemical, Cellular, and Histopathological Alterations. *Frontiers in Pharmacology* [Internet]. 2015;6:268. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC4637408/>
171. Ferreira GK, Rezin GT, Cardoso MR, Gonçalves CL, Borges LS, Vieira JS, et al. Brain energy metabolism is increased by chronic administration of bupropion. *Acta Neuropsychiatrica* [Internet]. 2012;24(2):115–21. Available from: <https://www.cambridge.org/core/article/brain-energy-metabolism-is-increased-by-chronic-administration-of-bupropion/7924065BDD149D8B83CDDE580259C9A7>
172. Abelaira HM, Réus GZ, Ribeiro KF, Zappellini G, Ferreira GK, Gomes LM, et al. Effects of acute and chronic treatment elicited by lamotrigine on behavior, energy metabolism, neurotrophins and signaling cascades in rats. *Neurochemistry International* [Internet]. 2011 Dec 1;59(8):1163–74. Available from: <http://www.sciencedirect.com/science/article/pii/S0197018611003421>
173. Della FP, Abelaira HM, Réus GZ, Ribeiro KF, Antunes AR, Scaini G, et al. Tianeptine treatment induces antidepressive-like effects and alters BDNF and energy metabolism in the brain of rats. *Behavioural Brain Research* [Internet]. 2012 Aug 1;233(2):526–35. Available from: <http://www.sciencedirect.com/science/article/pii/S0166432812003907>
174. Ignácio ZM, Réus G, Abelaira H, E. Titus S, S. Carlessi A, R. da Luz J, et al. Acute and Chronic Treatments with Quetiapine Increase Mitochondrial Respiratory Chain Complex Activity in the Rat Brain. Vol. 12. 2015.
175. Rezin GT, Gonçalves CL, Daufenbach JF, Fraga DB, Santos PM, Ferreira GK, et al. Acute administration of ketamine reverses the inhibition of mitochondrial respiratory chain induced by chronic mild stress. *Brain Research Bulletin* [Internet]. 2009 Aug 14;79(6):418–21. Available from: <http://www.sciencedirect.com/science/article/pii/S0361923009000963>
176. Della FP, Abelaira HM, Réus GZ, dos Santos MAB, Tomaz DB, Antunes AR, et al. Treatment with tianeptine induces antidepressive-like effects and alters the neurotrophin levels, mitochondrial respiratory chain and cycle Krebs enzymes in the brain of maternally deprived

- adult rats. *Metabolic Brain Disease* [Internet]. 2013 Mar 1;28(1):93–105. Available from: <https://doi.org/10.1007/s11011-012-9375-x>
177. Scaini G, Rochi N, Morais MOS, Maggi DD, De-Nês BT, Quevedo J, et al. In vitro effect of antipsychotics on brain energy metabolism parameters in the brain of rats. *Acta Neuropsychiatrica* [Internet]. 2013;25(1):18–26. Available from: <https://www.cambridge.org/core/article/in-vitro-effect-of-antipsychotics-on-brain-energy-metabolism-parameters-in-the-brain-of-rats/D9A7540ADE8D3625CF5563E69ABFEE22>
  178. Przedborski Serge, Jackson-Lewis Vernice, Fahn Stanley. Antiparkinsonian therapies and brain mitochondrial complex I activity. *Movement Disorders* [Internet]. 1995 May [cited 2018 May 20];10(3):312–7. Available from: <https://doi.org/10.1002/mds.870100314>
  179. Mohamed TM, Ghaffar HMA, El Husseiny RM. Effects of tramadol, clonazepam, and their combination on brain mitochondrial complexes. *Toxicol Ind Health* [Internet]. 2013 Jul 10 [cited 2018 May 11];31(12):1325–33. Available from: <https://doi.org/10.1177/0748233713491814>
  180. van der Kooij MA, Hollis F, Lozano L, Zalachoras I, Abad S, Zanoletti O, et al. Diazepam actions in the VTA enhance social dominance and mitochondrial function in the nucleus accumbens by activation of dopamine D1 receptors. *Molecular Psychiatry* [Internet]. 2018;23(3):569–78. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC5822450/>
  181. Kudin A, Debska-Vielhaber G, Vielhaber S, Elger C, Kunz W. The Mechanism of Neuroprotection by Topiramate in an Animal Model of Epilepsy. *Epilepsia* [Internet]. 2004 Nov 29 [cited 2018 May 17];45(12):1478–87. Available from: <https://doi.org/10.1111/j.0013-9580.2004.13504.x>
  182. Kumar P, Kumar A. Protective effect of rivastigmine against 3-nitropropionic acid-induced Huntington's disease like symptoms: Possible behavioural, biochemical and cellular alterations. *European Journal of Pharmacology* [Internet]. 2009 Aug 1;615(1):91–101. Available from: <http://www.sciencedirect.com/science/article/pii/S001429990900418X>
  183. Saravanan KS, Sindhu KM, Senthilkumar KS, Mohanakumar KP. 1-deprenyl protects against rotenone-induced, oxidative stress-mediated dopaminergic neurodegeneration in rats. *Neurochemistry International* [Internet]. 2006 Jul 1;49(1):28–40. Available from: <http://www.sciencedirect.com/science/article/pii/S0197018606000234>

## Figures

**Fig. 1. Forest plot: Drugs.** Multivariate estimates of the effects (SMD, 95% CI, p-values) of antidepressants, antipsychotics, anxiolytics, mood stabilizers, stimulants, antidementia drugs and antiparkinsonian drugs on complex I and IV enzyme activities. Values lower than 0 indicate that treated animals had lower levels than controls, and vice versa for values greater than 0; the dashed vertical line at SMD = 0 indicates no effect. The size of the filled circles for each estimated SMD is proportional to the weight of the studies.

**Fig. 2. Forest plot: Mechanism of action.** Multivariate estimates of the effects (SMD, 95% CI, p-values) on complex I and IV enzyme activities of mechanisms of action of antidepressants, antipsychotics, anxiolytics, mood stabilizers, stimulants, antidementia drugs and antiparkinsonian drugs. Values lower than 0 indicate that animals treated had lower levels than controls, and vice versa for values greater than 0; the dashed vertical line at SMD = 0 indicates no effect. The size of the filled circles for each estimated SMD is proportional to the weight of the studies.

Antipsychotics (APs), antiepileptic drugs (AEGs), cholinesterase inhibitors (ChEIs), dopamine agonists (DAGs), dopamine reuptake inhibitors (DRIs), gamma-aminobutyric acid (GABA)A receptor positive allosteric modulators (PAMs), monoamine oxidase inhibitors (MAOIs), muscarinic M1 agonists (M1 AGs), N-methyl-D-aspartate receptor antagonists (NMDA ANTs), noradrenergic and specific serotonergic antidepressants (NaSSAs), norepinephrine-dopamine disinhibitors (NDDIs), norepinephrine-dopamine reuptake inhibitors (NDRIs), norepinephrine reuptake inhibitors (NRIs), reversible inhibitors of monoamine oxidase A (RIMAs), serotonin antagonist and reuptake inhibitors (SARIs), serotonin-norepinephrine reuptake inhibitors (SNRIs), selective serotonin reuptake enhancers (SSREs), selective serotonin reuptake inhibitors (SSRIs).

**Fig. 3. Forest Plot: Receptor affinity.** Multivariate estimates of the effects (SMD, 95% CI, p-values) on complex I and IV enzyme activities by receptor affinity of antidepressants, antipsychotics, anxiolytics, mood stabilizers, stimulants, antidementia drugs and antiparkinsonian drugs. Values lower than 0 indicate that animals treated had lower levels than controls, and vice versa for values greater than 0; the dashed vertical line at SMD = 0 indicates no effect. The size of the filled circles for each estimated SMD is proportional to the weight of the studies. Type of receptor binding, i.e., agonism

(AG) or antagonism (ANT), is indicated. See **Fig. S1, Dataset S2** for included drugs and affinity profiles.

Adrenergic ( $\alpha,\beta$ ), dopaminergic (D), GABAergic (GABA-A), glutaminergic (NMDA, AMPA), histaminergic (H), muscarinic (M), nicotinic (nACh), opioid (OP), serotonergic (5-HT), sigma ( $\sigma$ ). DAT, NET and SERT correspond to the abilities of the drugs to inhibit the reuptake of dopamine, norepinephrine and serotonin, respectively.

**Tab. 1. Eligible studies.** 68 studies were eligible for the meta-analyses covering 53 drugs. Mechanisms of actions are indicated <sup>56</sup>. Some studies reported more than one drug. The detailed data extracted from these studies are provided in **Dataset S1 & S2**.

Antipsychotics (APs), antiepileptic drugs (AEGs), cholinesterase inhibitors (ChEIs), dopamine agonists (DAGs), dopamine reuptake inhibitors (DRIs), gamma-aminobutyric acid (GABA)A receptor positive allosteric modulators (PAMs), monoamine oxidase inhibitors (MAOIs), muscarinic M1 agonists (M1 AGs), N-methyl-D-aspartate receptor antagonists (NMDA ANTs), noradrenergic and specific serotonergic antidepressants (NaSSAs), norepinephrine-dopamine disinhibitors (NDDIs), norepinephrine-dopamine reuptake inhibitors (NDRIs), norepinephrine reuptake inhibitors (NRIs), reversible inhibitors of monoamine oxidase A (RIMAs), serotonin antagonist and reuptake inhibitors (SARIs), serotonin-norepinephrine reuptake inhibitors (SNRIs), selective serotonin reuptake enhancers (SSREs), selective serotonin reuptake inhibitors (SSRIs).

Antidepressants	Complex	Mechanism	Author
Agomelatine	I, IV	NDDI	Gupta 2014 <sup>85</sup> , Kumar 2015 <sup>86</sup> , Singh 2015 <sup>170</sup> , de Mello 2015 <sup>84</sup>
Amitriptyline	I, IV	NRI	González-Pardo 2008 <sup>69</sup> , Hroudová 2010 <sup>16</sup>
Bupropion	I, IV	NDRI	Ferreira 2012 <sup>171</sup>
Citalopram	I, IV	SSRI	Hroudová 2010 <sup>16</sup>
Desipramine	I, IV	NRI	Hroudová 2010 <sup>16</sup> , Villa 2017 <sup>64</sup>
Escitalopram	I, IV	SSRI	Gonçalves 2012 <sup>59</sup>
Fluoxetine	I, IV	SSRI	Adzic 2013 <sup>60</sup> , Padilla 2011 <sup>62</sup> , Shumake 2010 <sup>63</sup> , Villa 2017 <sup>64</sup>
Fluvoxamine	I, IV	SSRI	Ferreira 2014 <sup>61</sup>
Imipramine	I, IV	SNRI	Abelaira 2011 <sup>172</sup> , Della 2012 <sup>173</sup> , Hroudová 2010 <sup>16</sup> , Ignácio 2015 <sup>174</sup> , Kumar 2010 <sup>68</sup> , Réus 2012 <sup>148</sup>
Ketamine	I, IV	NMDA ANT	Rezin 2009 <sup>175</sup> , Rezin 2010 <sup>94</sup> , Réus 2015 <sup>93</sup> , Venâncio 2015 <sup>95</sup> , de Oliveira 2011 <sup>92</sup>
Mirtazapine	I, IV	NaSSA	Hroudová 2010 <sup>16</sup>
Moclobemide	I, IV	RIMA	Hroudová 2010 <sup>16</sup>
Nortriptyline	I, IV	NRI	Scaini 2011 <sup>66</sup>
Paroxetine	I, IV	SSRI	Garabadu 2015 <sup>65</sup> , Scaini 2011 <sup>66</sup>
Sertraline	I, IV	SSRI	Gaur 2010 <sup>67</sup> , Kumar 2010 <sup>68</sup> , Sharma 2016 <sup>163</sup>
Tianeptine	I, IV	SSRE	Della 2012 <sup>173</sup> , Della 2013 <sup>176</sup> , Hroudová 2010 <sup>16</sup>
Trazodone	I, IV	SARI	Kumar 2010 <sup>68</sup>
Venlafaxine	I, IV	SNRI	Hroudová 2010 <sup>16</sup> , Kumar 2010 <sup>68</sup> , Scaini 2011 <sup>66</sup> , Sharma 2016 <sup>163</sup>
Antipsychotics	Complex	Mechanism	Author
Aripiprazole	I, IV	Atypical AP	Scaini 2013 <sup>177</sup> , Streck 2007 <sup>104</sup>
Chlorpromazine	I, IV	Typical AP	Balijepalli 1999 <sup>98</sup> , Burkhardt 1993 <sup>101</sup>
Clozapine	I, IV	Atypical AP	Balijepalli 1999 <sup>98</sup> , Balijepalli 2001 <sup>99</sup> , Burkhardt 1993 <sup>101</sup> , Prince 1997 <sup>102</sup> , Prince 1998 <sup>103</sup> , Przedborski 1995 <sup>178</sup> , Scaini 2013 <sup>177</sup> , Streck 2007 <sup>104</sup>
Fluphenazine	I, IV	Typical AP	Balijepalli 1999 <sup>98</sup> , Prince 1997 <sup>102</sup> , Prince 1998 <sup>103</sup>
Haloperidol	I, IV	Typical AP	Balijepalli 1999 <sup>98</sup> , Balijepalli 2001 <sup>99</sup> , Barrientos 1998 <sup>100</sup> , Burkhardt 1993 <sup>101</sup> , Prince 1997 <sup>102</sup> , Prince 1998 <sup>103</sup> , Streck 2007 <sup>104</sup>
Molindone	I	Typical AP	Przedborski 1995 <sup>178</sup>
Olanzapine	I, IV	Atypical AP	Hroudová 2010 <sup>16</sup> , Scaini 2013 <sup>177</sup> , Streck 2007 <sup>104</sup>
Quetiapine	I, IV	Atypical AP	Ignácio 2015 <sup>174</sup>
Risperidone	I, IV	Atypical AP	Balijepalli 1999 <sup>98</sup> , Balijepalli 2001 <sup>99</sup> , Garabadu 2015 <sup>65</sup>

Thiothixene	I, IV	Typical AP	Burkhardt 1993 <sup>101</sup>
<b>Anxiolytics</b>	<b>Complex</b>	<b>Mechanism</b>	<b>Author</b>
Clonazepam	I, IV	GABA-A PAM	Mohamed 2013 <sup>179</sup>
Diazepam	I, IV	GABA-A PAM	Kumar 2014 <sup>132</sup> , van der Kooij 2018 <sup>180</sup>
<b>Mood stabilizers</b>	<b>Complex</b>	<b>Mechanism</b>	<b>Author</b>
Gabapentin	I, IV	AEG	Kumar 2012 <sup>131</sup> , Kumar 2014 <sup>132</sup>
Lamotrigine	I, IV	AEG	Abelaira 2011 <sup>172</sup> , Kumar 2012 <sup>131</sup>
Lithium	I, IV	Lithium	Bachmann 2009 <sup>119</sup> , Feier 2013 <sup>120</sup> , Hroudová 2010 <sup>16</sup> , Kim 2016 <sup>121</sup> , Lambert 1999 <sup>122</sup> , Tan 2012 <sup>123</sup> , Valvassori 2010 <sup>124</sup>
Topiramate	I, IV	AEG	Kudin 2004 <sup>181</sup>
Valproic acid	I, IV	AEG	Bachmann 2009 <sup>119</sup> , Feier 2013 <sup>120</sup> , Hroudová 2010 <sup>16</sup> , Mishra 2014 <sup>140</sup> , Valvassori 2010 <sup>124</sup>
<b>Stimulants</b>	<b>Complex</b>	<b>Mechanism</b>	<b>Author</b>
Amphetamine	I	DRI	Tan 2012 <sup>123</sup>
Mazindol	I, IV	DRI	Gonçalves 2014 <sup>138</sup>
Methamphetamine	I, IV	DRI	Brown 2005 <sup>135</sup> , Killinger 2014 <sup>139</sup> , Thrash 2010 <sup>141</sup> , Thrash-Williams 2013 <sup>142</sup> , Thrash-Williams 2016 <sup>143</sup>
Methylphenidate	I, IV	DRI	Fagundes 2007 <sup>136</sup> , Fagundes 2010 <sup>137</sup>
<b>Antidementia</b>	<b>Complex</b>	<b>Mechanism</b>	<b>Author</b>
7-MEOTA	I, IV	Unknown	Singh 2017 <sup>149</sup>
Donepezil	I, IV	ChEI	Singh 2015 <sup>170</sup> , Singh 2017 <sup>149</sup>
Galantamine	I, IV	ChEI	Singh 2015b <sup>87</sup> , Singh 2017 <sup>149</sup>
Latrepirdine	I, IV	Unknown	Singh 2017 <sup>149</sup>
Memantine	I, IV	NMDA ANT	Kumar 2016 <sup>147</sup> , Réus 2012 <sup>148</sup> , Singh 2017 <sup>149</sup>
Piracetam	I, IV	Unknown	Singh 2017 <sup>149</sup>
Rivastigmine	I, IV	ChEI	Kumar 2009 <sup>182</sup> , Singh 2017 <sup>149</sup>
<b>Antiparkinsonian</b>	<b>Complex</b>	<b>Mechanism</b>	<b>Author</b>
Amantadine	I	Unknown	Thrash-Williams 2013 <sup>142</sup>
Bromocriptine	I	DAG	Przedborski 1995 <sup>178</sup>
Levodopa	I, IV	DAG	Abdin 2008 <sup>160</sup> , Calabrese 2007 <sup>161</sup> , Dixit 2013 <sup>159</sup> , Przedborski 1993 <sup>162</sup> , Sharma 2016 <sup>163</sup>
Minocycline	I	Unknown	Dixit 2013 <sup>159</sup>
Pergolide	I	DAG	Przedborski 1995 <sup>178</sup>
Selegiline	I, IV	MAOI	Czerniczyniec 2006 <sup>164</sup> , Przedborski 1995 <sup>178</sup> , Saravanan 2006 <sup>183</sup>
Trihexyphenidyl	I	M1 AG	Przedborski 1995 <sup>178</sup>

**Tab. 2. Heterogeneity and Inconsistency.** Significant Q statistics indicate the existence of heterogeneity. A  $I^2$  value of 0% indicates no observed inconsistency, whereas larger values shows increasing inconsistency. **Egger's regression test for publication bias.** The existence of potential publication bias is indicated by p-values < 0.05. df = degrees of freedom.

<b>Heterogeneity and Inconsistency</b>				
		<b>Q</b>	<b>p-value</b>	<b><math>I^2</math> (%)</b>
<b>Complex I</b>	<b>Antidepressants</b>	682.00	0.000	86%
	<b>Antipsychotics</b>	533.87	0.000	87%
	<b>Anxiolytics</b>	34.99	0.000	89%
	<b>Mood stabilizers</b>	131.09	0.000	85%
	<b>Stimulants</b>	45.93	0.434	11%
	<b>Antidementia drugs</b>	122.33	0.000	71%
	<b>Antiparkinsonian drugs</b>	83.39	0.000	84%
<b>Complex IV</b>	<b>Antidepressants</b>	707.35	0.000	71%
	<b>Antipsychotics</b>	661.47	0.000	76%
	<b>Anxiolytics</b>	4.57	0.102	87%
	<b>Mood stabilizers</b>	339.63	0.000	80%
	<b>Stimulants</b>	111.71	0.000	53%
	<b>Antidementia drugs</b>	57.39	0.000	63%
	<b>Antiparkinsonian drugs</b>	5.58	0.589	34%
<b>Egger's regression</b>				
		<b>z-value</b>	<b>df</b>	<b>p-value</b>
<b>Complex I</b>	<b>Antidepressants</b>	1.74	22	0.095
	<b>Antipsychotics</b>	-1.36	8	0.211
	<b>Anxiolytics</b>	0.49	2	0.671
	<b>Mood stabilizers</b>	1.54	8	0.161
	<b>Stimulants</b>	-0.13	7	0.902
	<b>Antidementia drugs</b>	-1.16	4	0.311
	<b>Antiparkinsonian drugs</b>	1.23	7	0.258
<b>Complex IV</b>	<b>Antidepressants</b>	0.44	24	0.665
	<b>Antipsychotics</b>	0.23	7	0.822
	<b>Anxiolytics</b>	3.73	1	0.167
	<b>Mood stabilizers</b>	1.10	8	0.302
	<b>Stimulants</b>	0.64	4	0.558
	<b>Antidementia drugs</b>	-0.07	3	0.949
	<b>Antiparkinsonian drugs</b>	0.97	1	0.511



Figure1  
Click here to download high resolution image

This preprint research paper has not been peer reviewed. Electronic copy available at: <https://ssrn.com/abstract=3235666>

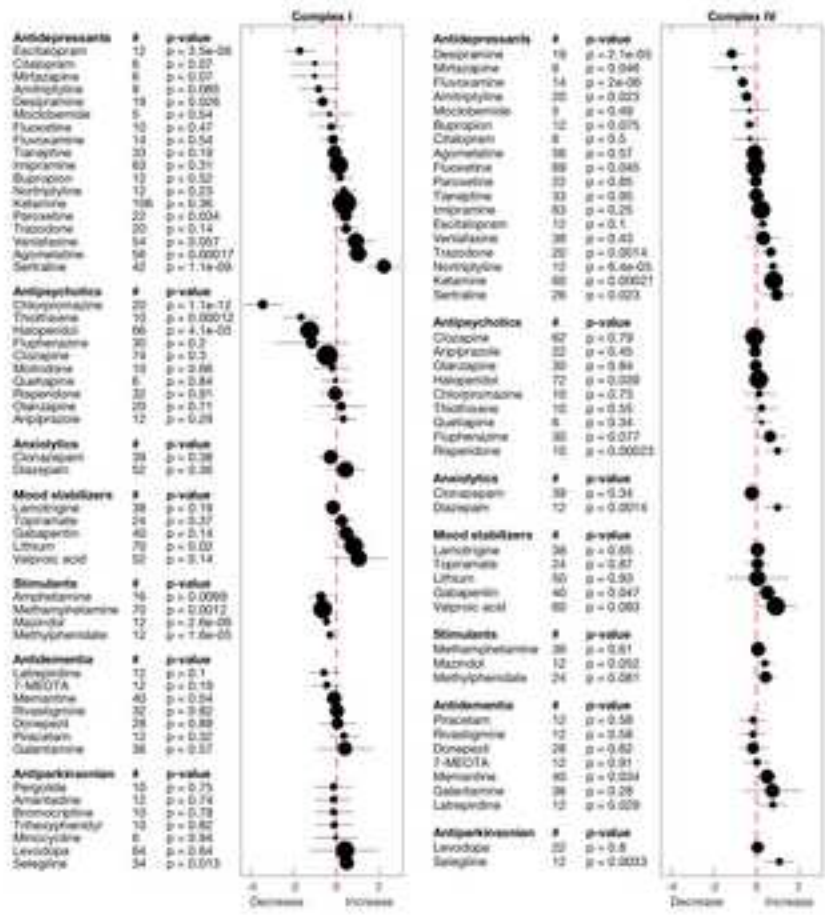


Figure2  
Click here to download high resolution image

This preprint research paper has not been peer reviewed. Electronic copy available at: <https://ssrn.com/abstract=3235666>

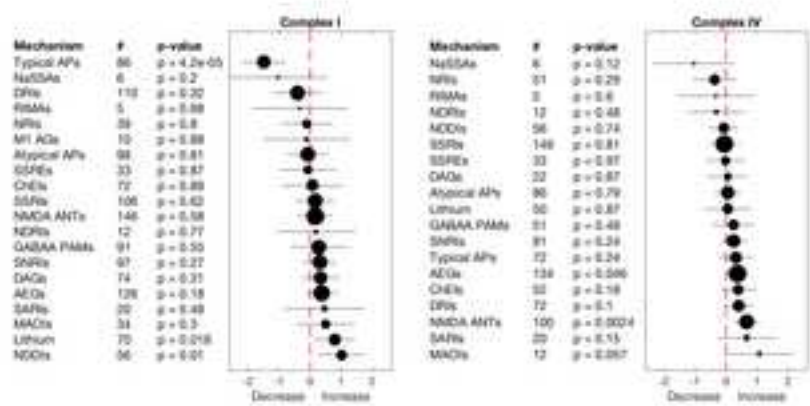
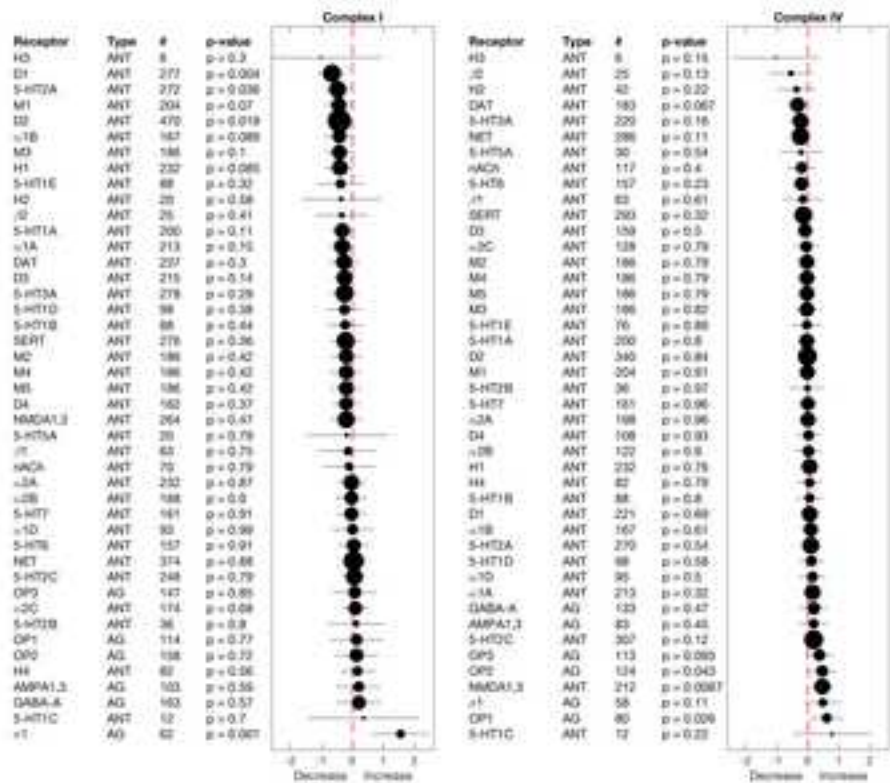


Figure3  
Click here to download high resolution image

This preprint research paper has not been peer reviewed. Electronic copy available at: <https://ssrn.com/abstract=3235666>



**Dataset 1**  
[Click here to download Necessary Additional Data: DatasetS1.txt](#)

**Dataset 2**  
**[Click here to download Necessary Additional Data: DatasetS2.txt](#)**

## Supplementary Materials

[Click here to download Necessary Additional Data: SupplementaryMaterials.docx](#)